

NSTITUTE OF SCIENCE, TECHNOLOGY & ADVANCED STUDIES (VISTAS) (Deemed is be University East, six 3 of the UGC Act, 1956) PALLAVARAM - CHENNAI ACCREDITED BY NAAC WITH 'A' GRADE Marching Beyond 30 Years Successfully INSTITUTION WITH UGC 12B STATUS

UNDERGRADUATE DEGREE PROGRAMME

B.Sc., Microbiology Three Years / B.Sc., (Hons) Microbiology Four Years

CURRICULUM & SYLLABUS

REGULATION 2024

Choice Based Credit System (CBCS)

&

Learning Outcomes Based Curriculum Framework (LOCF)

Effective from the Academic Year 2024 -2025

Department of Microbiology School of Life Sciences



INSTITUTE OF SCIENCE, TECHNOLOGY & ADVANCED STUDIES (VISTAS) (Deemed is be University End. us/3 of the UGC Act, 1956) PALLAWARAM - CHENNAI ACCREDITED BY NAAC WITH 'A' GRADE Marching Beyond 30 Years Successfully INSTITUTION WITH UGC 12B STATUS

DEPARTMENT OF MICROBIOLOGY

VISION OF THE DEPARTMENT

To produce graduates with relevant education descriptors and hands-on skills in microbiology and related areas of life sciences plus holistic development of individuals that makes them responsible citizens of society.

	MISSION OF THE DEPARTMENT
M1	Imparting relevant knowledge and creating an atmosphere to develop innovative and critical thinking.
M2	Skill enhancement through hands-on training and value-added courses plus add on courses.
M3	Sustained focus on original high-quality research encouraging scientific thinking and approach.
M4	Creating an environment for holistic development of individuals with emphasis on spirit of integrity, equity, professional ethics and social harmony through the exposure and participation in co-curricular, extracurricular and extension activities.

	PROGRAMME EDUCATIONAL OUTCOMES (PEO)									
	To provide the graduates with knowledge in microbiology and an overview of the									
PEO1	processes that employ or deal with microbes that enables them to handle the safe and									
	efficient use of microbiological applications with development of competence on par									
	with global standards and helps the graduates for life-long learning.									
	To prepare graduates by imparting skills to use technological developments related									
PEO2	to current and advanced areas involving molecular diagnostics, immunotechnology, mass cultivation of microbes, downstream processing and nanotechnology with									
PEO2										
	scope for upskilling in all potential future technologies so as to contribute effectively									
	for Research & Development leading to patenting and publishing.									
	To train graduates to choose a decent career option either as Entrepreneur or having									
PEO3	a high degree of employability; or pursue higher education - by empowering									
TEOS	students with basic interpersonal skills, ability to handle critical situations allowing									
	them to be good team members as well as training to excel in competitive									
	examinations.									
PEO4	To impart a strong sense of social responsibility with awareness of professional and									
	societal ethical values and scope to develop leadership capabilities.									
PEO5	To establish an environment that accentuates the requirement to fulfil life-long									
	learning for the overall development of self and society at large.									

	PROGRAMME OUTCOMES (PO)
PO1	Microbiology knowledge: Graduates will acquire microbiology specific knowledge
PUI	including molecular biology, immunology and rDNA technology coupled with hands-
	on skills and leadership skills for a successful career.
PO2	Problem analysis: Graduates will be able to analyse, solve and troubleshoot
	problems in implementation of microbiological protocols.
PO3	Design/development of solutions: Graduates will develop creative thinking and
	cooperate with each other to solve problems in the field of microbiology.
PO4	Conduct investigations of complex problems: Graduates will acquire practical skills
104	

	- which help in planning and designing protocols to validate hypothesis and execute									
	experimental techniques independently as well as assimilate, analyse and interpret									
	subsequent data.									
	Modern tool usage and communication: Graduates will effectively be able to									
PO5	manage resources and time using ICT and computer enabled devices and accomplish									
	ability to understand and communicate all ideas effectively.									
	Environment sustainability and Ethics: Graduates will get adequate knowledge to									
PO6	use information and implement solutions for environmental protection and									
	remediation. Graduates will be aware of their role and responsibility in handling and									
	use of microbes including genetically modified microorganisms.									
PO7	Lifelong learning: Graduates will carry on to learn and adapt in a world of constantly									
	evolving technology.									

	PROGRAMME SPECIFIC OUTCOMES (PSO)
PSO1	Microbiology skills: The ability to understand the basic concepts related to the
rsoi	relevant fields of microbiology which will enable them to analyse and develop
	solutions to microbiology related problems.
PSO2	Microbiology related employability skills: The ability to use the acquired hands-on
FSU2	skills in microbiology, molecular biology, immunology, medical microbiology and
	screening for useful biomolecules within employment areas.
	Successful Career and Entrepreneurship: The ability to gainfully become an
D CO2	entrepreneur by using microorganisms to produce biofertilizers, mushrooms and
PSO3	pharmaceutically important biomolecules as well as using practical hands-on training
	to become employed in diagnostic, industrial, pharmaceutical, food and research and
	development laboratories.

BOARD OF STUDIES

List of Members

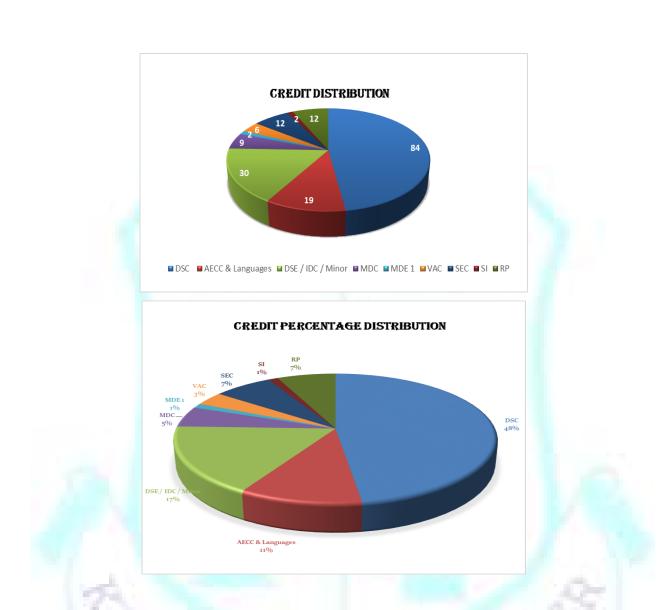
Department of Microbiology

S. No	Name & Designation	Address	Role
1	Dr.G.Gayathri Associate Professor and Head I/c Department of Microbiology, VISTAS. Chennai.	Associate Professor and Head I/c Department of Microbiology, VISTAS. Chennai.	Chairperson
2	Dr. A.K.Kathireshan Professor & Director, School of Life Sciences, VISTAS, Chennai.	Professor & Director School of Life Sciences, VISTAS. Chennai.	Member
3	Dr.Suresh Dhanaraj Associate Professor Department of Microbiology, VISTAS. Chennai.	Associate Professor Department of Microbiology, VISTAS. Chennai.	Member
4	Dr.S.B.Prabha Assistant Professor Department of Microbiology, VISTAS. Chennai.	Assistant Professor Department of Microbiology, VISTAS. Chennai.	Member
5	Dr.S.V.Rajyoganandh Assistant Professor Department of Microbiology, VISTAS. Chennai.	Assistant Professor Department of Microbiology, VISTAS. Chennai.	Member
6	Dr. K.Mahalakshmi Professor and Head	Director – Research lab for Oral and Systemic Health, Department of Microbiology, Sree Balaji Dental College and Hospital, Pallikaranai, Bharath University, Chennai.	Academic Expert (External Member)
7	Dr.Babu Sarangan Managing Director, Mahathi Biotech.	Mahathi Biotech K.K Nagar, Chennai – 78.	Industrial Expert (External Member)
8	Thiru Siraj Kareem Managing Director Inlead Management	Inlead Management Services, Mangadu, Chennai.	Alumini (External Member)

CREDIT DISTRIBUTION

B.Sc., (Hons) in Microbiology Minimum credits to be earned: 176 B.Sc., Microbiology Minimum credits to be earned: 132

Component	I Sem	II Sem	III Sem	IV Sem	V Sem	VI Sem	3 Yrs. Total Credits	VII Sem	VIII Sem	4 Yrs Total Credits
DSC	8	8	8	8	12	16	60	12	12	84
AECC & Languages	4	4	4	7	-	-	19	-	-	19
DSE / IDC / Minor	4	4	3	4	4	3	22	4	4	30
MDC	3	3	3	-	-	-	9	-	-	9
MDE 1	-	-	2	-	-	-	2	-	-	2
VAC	1	2	-	1	2	-	6	-	-	6
SEC	2	2	2	2	2	2	12	-		12
SI	-	-	1	-	1	-	2	-	- 1	2
RP	-	-	-	-	-	-	-	6	6	12
Total Credits	22	23	23	22	21	21	132	22	22	176
		-	E	G	EI	5)	-	1		



ABBREVIATIONS

DSC	Disciplinary Specific Core
AECC	Ability Enhancement Compulsory Courses
DSE	Disciplinary Specific Elective
IDC	Interdisciplinary / Minor Courses
MDC	Multidisciplinary Courses
VAC	Value Added Courses
SEC	Skill Enhancement Courses
SI	Summer Internship
RP	Research Project

CURRICULUM STURCTURE

B.Sc., Microbiology Three Years

BSc., (Hons) in Microbiology Four Years

									redits: 1				
		B.Sc., Microbiol	ogy (Hons) Mini	mum	Credi	ts to be	earned	:176			
	B.Sc., Microbiology Minimum Credits to be earned: 132 SEMESTER 1												
	<u> </u>			Ho	urs/W	'eek		Ma	ximum 1	Marks			
Category	Code	Course	L	Т	Р	0	С	CIA	SEE	Total			
LANG 1	24LTAM11/2 4LHIN11/ 24LFRE11	Tamil I / Hindi I/ French I	2	0	0	1	2	40	60	100			
ENG 1	24LENG11	English I	2	0	0	1	2	40	60	100			
DSC 1	24CBMB11	Basic Techniques in Microbiology	3	0	0	2	3	40	60	100			
DSC 2	24CBMB12	General Microbiology	4	0	0	2	4	40	60	100			
MDC 1	24MBMB11	Microbiology of Air and Water	3	0	0	2	3	40	60	100			
DSE 1/ IDC 1 / Minor 1	24DBMB1-	Discipline Specific Elective - I	4	0	0	2	4	40	60	100			
DSC 1 (Lab)	24PBMB11	Practical I - Practical in Basic Microbiological Techniques	0	0	2	1	1	40	60	100			
VAC 1	24DVAC11	Universal Human Values	1	0	0	1	1	1	100	100			
SEC 1	24SSKU11	Soft Skills I	2	0	0	1	2	40	60	100			
SEC 2		Orientation programme / Industrial Visit	-	-	-	-	-	-	-	-			
			21	-	2	-	22	-	-	-			

		SEME	STE	R 2						
Category	Code	Course	L	Τ	Р	0	С	CIA	SEE	Total
LANG 2	24LTAM21/ 24LHIN21/ 24LFRE21	Tamil II / Hindi II / French II	2	0	0	1	2	40	60	100
ENG 2	24LENG21	English II	2	0	0	1	2	40	60	100
DSC 3	24CBMB21	Microbial Genetics and Molecular Biology	3	0	0	2	3	40	60	100
DSC 4	24CBMB22	Industrial Microbiology	3	0	0	2	3	40	60	100
MDC 2	24MBMB21	Bioinstrumentation	3	0	0	2	3	40	60	100
DSE 2 / IDC 2 / Minor 2	24DBMB2-	Discipline Specific Elective - II	4	0	0	2	4	40	60	100
DSC 3 (Lab)	24PBMB21	Practical II – Practical in Molecular biology	0	0	2	1	1	40	60	100
DSC 4 (Lab)	24PBMB22	Practical III – Practical in Industrial Microbiology	0	0	2	1	1	40	60	100
VAC 2	24DVAC21	Communication Skills	2	0	0	1	2	40	60	100
SEC 3	24SSKU21	Soft Skills II	2	0	0	1	2	40	60	100
	100	ar.	21	-	4	C	23		-	-

CIA - Continuous Internal Assessment SEE - Semester End Examination *L – Lecture, *T- Tutorial, *P- Practical, *O - Outside the class effort / self-study

		SEM	ESTE	CR 3						
Category	Code	Course	L	Т	Р	0	C	CIA	SEE	Total
LANG 3	24LTAM31/ 24LHIN31/ 24LFRE31	Tamil III / Hindi III / French III	2	0	0	1	2	40	60	100
ENG 3	24LENG31	English III	2	0	0	1	2	40	60	100
DSC 5	24CBMB31	Basic Concepts in Immunology	3	0	0	2	3	40	60	100
DSC 6	24CBMB32	Environmental Microbiology	3	0	0	2	3	40	60	100
MDC 3	24MBMB31	Microbial Biotechnology	3	0	0	2	3	40	60	100
DSE 3 / IDC 3 / Minor 3	24DBMB3-	Discipline Specific Elective - II	3	0	0	2	3	40	60	100
DSC 5 (Lab)	24PBMB31	Practical IV – Practical in Immunology	0	0	2	1	1	40	60	100
DSC 6 (Lab)	24PBMB32	Practical V – Practical in Environmental Microbiology	0	0	2	1	1	40	60	100
MDE 1	30	Indian Knowledge System	1	1	0	1	2	40	60	100
SEC 4	24SSKU31	Soft Skills III	2	0	0	1	2	40	60	100
SI 1	24IBMB31	Internship I	0	0	2	1	1	-	100	100
			19	1	6	-	23	-	-	-

CIA - Continuous Internal Assessment SEE - Semester End Examination

SEMESTER 4										
Category	Code	Course	L	Т	P	0	C	CIA	SEE	Total
LANG 4	24LTAM41/ 24LHIN41/ 24LFRE41	Tamil IV / Hindi IV / French IV	2	0	0	1	2	40	60	100
ENG 4	24LENG41	English IV	2	0	0	1	2	40	60	100
AECC 1	24EVS031	Environmental Studies	3	0	0	2	3	40	60	100
DSC 7	24CBMB41	Medical Mycology and Parasitology	3	0	0	2	3	40	60	100
DSC 8	24CBMB42	Medical Bacteriology	3	0	0	2	3	40	60	100
DSE 4 / IDC 4 / Minor 4	24DBMB4-	Discipline Specific Elective - IV	4	0	0	2	4	40	60	100
DSC 7 (Lab)	24PBMB41	Practical VI – Practical in Mycology and Parasitology	0	0	2	1	1	40	60	100
DSC 8 (Lab)	24PBMB42	Practical VII – Practical in Bacteriology	0	0	2	1	1	40	60	100
SEC 5	24SBMB41	Industry Oriented Employability skills	1	0	2	1	2	40	60	100
VAC 3	24DVAC41	Yoga Education / NSS / NCC	0	0	2		1	2	100	100
SEC 6		In-plant Training/ Industrial Tour/ Summer Term	FE	15	2	-	-	-	-	-
			18	-	8	-	22	-	-	-

CIA - Continuous Internal Assessment SEE - Semester End Examination

		SEN	IEST	ER 5						
Category	Code	Course	L	Т	P	0	C	CIA	SEE	Total
DSC 9	24CBMB51	Soil & Agricultural Microbiology	3	0	0	2	3	40	60	100
DSC 10	24CBMB52	Food Microbiology	3	0	0	2	3	40	60	100
DSC 11	24CBMB53	Basics of Pharmaceutical Microbiology	4	0	0	2	4	40	60	100
DSE 5 / IDC 5 / Minor 5	24DBMB5-	Discipline Specific Elective - V	4	0	0	2	4	40	60	100
DSC 9 (Lab)	24PBMB51	Practical VIII – Practical in Soil and Agricultural Microbiology	0	0	2	1	1	40	60	100
DSC 10 (Lab)	24PBMB52	Practical IX - Practical in Food and Pharmaceutical Microbiology	0	0	2	1	1	40	60	100
SEC 7	24SBMB51	Entrepreneurial Development	2	0	0	1	2	40	60	100
VAC 4	24DVAC51	Spirulina Cultivation	2	0	0	1	2	40	60	100
SI 2	24IBMB51	Internship II	0	0	2	1	1		100	100
SEC 8		Skill Enhancement Training / Student Club Activities/ Institution Innovation Council Activities		15	2		-	-	-	-
			18	-	6	-	21	-	-	-

		SEN	MEST	E R 6						
Category	Code	Course	L	Т	Р	0	С	CIA	SEE	Total
DSC 12	24CBMB61	Basics of Genetic Engineering	3	0	0	2	3	40	60	100
DSC 13	24CBMB62	Quality Assurance	4	0	0	2	4	40	60	100
DSC 14	24CBMB63	Biofertilizers Technology	4	0	0	2	4	40	60	100
DSC 15	24CBMB64	Mushroom Technology	4	0	0	2	4	40	60	100
DSE 6 / IDC 6 / Minor 6	24DBMB6-	Discipline Specific Elective - II	3	0	0	2	3	40	60	100
DSC 12 (Lab)	24PBMB61	Practical X – Practical in Genetic Engineering	0	0	2	1	1	40	60	100
SEC 9	24SBMB61	Mini Project	0	0	4	1	2	-	100	100
SEC 10	A	On Job Training / Apprenticeship / Startup	-	-	-	-	-	-	2	_
	de.		18	-	6	-	21	e <		-

CIA - Continuous Internal Assessment SEE - Semester End Examination

		SEME	STER	R 7						
Category	Code	Course	L	Τ	Р	0	C	CIA	SEE	Total
DSC 16	24CBMB71	Microbial	3	0	0	2	3	40	60	100
		Enzymology and								
		Metabolism								
DSC 17	24CBMB72	Microbes and	4	0	0	2	2 4 40	40	60	100
		Sustainable	1.000		1.1.1.1					
		Development				1.5				
DSC 18	24CBMB73	Research	4	0	0	2	4	40	60	100
		Methodology	1.1		16					
DSE 7 /		Discipline Specific	4	0	0	2	4	40	60	100
IDC 7 /	24DBMB7-	Elective - II								
Minor 7		Elective - II	-							
DSC 16	24PBMB71	Practical XI –	0	0	2	1	1	40	60	100
(Lab)		Practical in Microbial								
		Enzymology								
RP 1	24RBMB71	Research Project I	0	0	12	4	6	40	60	100
			15	-	14	-	22	-	-	-

CIA - Continuous Internal Assessment SEE - Semester End Examination

*L – Lecture, *T- Tutorial, *P- Practical, *O - Outside the class effort / self-study

		SEME	STEF	R 8						
Category	Code	Course	L	Τ	Р	0	C	CIA	SEE	Total
DSC 19	24CBMB81	Algal Biotechnology	3	0	0	2	3	40	60	100
DSC 20	24CBMB82	Marine Microbiology	3	0	0	2	3	40	60	100
DSC 21	24CBMB83	Microbial Entrepreneurship	4	0	0	2	4	40	60	100
DSE 8 / IDC 8 / Minor 8	24DBMB8-	Discipline Specific Elective - II	4	0	0	2	4	40	60	100
DSC 19 (Lab)	24PBMB81	Practical XII – Practical in Algal Biotechnology	0	0	2	1	1	40	60	100
DSC 20 (Lab)	24PBMB82	Practical XIII – Practical in Marine Microbiology	0	0	2	1	1	40	60	100
RP 2	24RBMB81	Research Project II	0	0	12	4	6	-	60	100
			14	-	16	-	22	-	-	-

CIA - Continuous Internal Assessment SEE - Semester End Examination

Category	Code	Course	L	Τ	Р	0	С
DSC 1	24CBMB11	Basic Techniques in Microbiology	3	0	0	2	3
DSC 2	24CBMB12	General Microbiology	4	0	0	2	4
DSC 1 (Lab)	24PBMB11	Practical I - Practical in Basic Microbiological Techniques	0	0	2	1	1
DSC 3	24CBMB21	Microbial Genetics and Molecular Biology	3	0	0	2	3
DSC 4	24CBMB22	Industrial Microbiology	3	0	0	2	3
DSC 3 (Lab)	24PBMB21	Practical II – Practical in Molecular biology	0	0	2	1	1
DSC 4 (Lab)	24PBMB22	Practical III – Practical in Industrial Microbiology	0	0	2	1	1
DSC 5	24CBMB31	Basic Concepts in Immunology	3	0	0	2	3
DSC 6	24CBMB32	Environmental Microbiology	3	0	0	2	3
DSC 5 (Lab)	24PBMB31	Practical IV – Practical in Immunology	0	0	2	1	1
DSC 6 (Lab)	24PBMB32	Practical V – Practical in Environmental Microbiology	0	0	2	1	1
DSC 7	24CBMB41	Medical Mycology and Parasitology	3	0	0	2	3
DSC 8	24CBMB42	Medical Bacteriology & Virology	3	0	0	2	3
DSC 7	24PBMB41	Practical VI –	0	0	2	1	1

DISCIPLINE SPECIFIC CORE COURSES

(Lab)		Practical in					
		Mycology and					
		Parasitology					
DECO		Practical VII –					
DSC 8	24PBMB42	Practical in	0	0	2	1	1
(Lab)		Bacteriology					
DCCA		Soil & Agricultural		0	0	-	2
DSC 9	24CBMB51	Microbiology	3	0	0	2	3
DSC 10	24CBMB52	Food Microbiology	3	0	0	2	3
		Basics of					
DSC 11	24CBMB53	Pharmaceutical	4	0	0	2	4
		Microbiology					
		Practical VIII –					
DSC 9		Practical in Soil and					
(Lab)	24PBMB51	Agricultural	0	0	2	1 1	1
Ì		Microbiology					
		Practical IX -					
DSC 10		Practical in Food and					1
(Lab)	24PBMB52	Pharmaceutical	0	0	2	1	1
, í	11.5	Microbiology					-
		Basics of Genetic					0
DSC 12	24CBMB61	Engineering	3	0	0	2	3
20010		Quality Assurance in					
DSC 13	24CBMB62	Microbiology	4	0	0	2	4
Dagas		Biofertilizers			0	-	
DSC 14	24CBMB63	Technology	4	0	0	2	4
		Mushroom					
DSC 15	24CBMB64	Technology	4	0	0	2	4
DOC 12		Practical X –					
DSC 12	24PBMB61	Practical in Genetic	0	0	2	1	1
(Lab)		Engineering					
Dage		Microbial				_	
DSC 16	24CBMB71	Enzymology and	3	0	0	2	3

		Metabolism					
DSC 17	24CBMB72	Microbes and Sustainable	4	0	0	2	4
DSC 18 DSC 16 (Lab) DSC 19		Development					
DSC 18	24CBMB73	Research	4	0	0	2	4
22010		Methodology					
DSC 16	17	Practical XI –	1				
(Lab)	24PBMB71	Practical in Microbial	0	0	2	1	1
	Y	Enzymology	\sim	2			
DSC 19	24CBMB81	Algal Biotechnology	3	0	0	2	3
DSC 20	24CBMB82	Marine Microbiology	3	0	0	2	3
DSC 21	24CBMB83	Microbial	4	0	0	2	4
	21000000	Entrepreneurship			Ŭ	_	'
DSC 19		Practical XII –					
	24PBMB81	Practical in Algal	0	0	2	1	1
		Biotechnology					
DSC 20		Practical XIII –					
	24PBMB82	Practical in Marine	0	0	2	1	1
` ´		Microbiology					

Category	Code	Course	L	Τ	Р	0	С
	24DBMB11	Microbial Diversity	4	0	0	2	4
DSE 1	24DBMB12	Microbial Physiology and Metabolism	4	0	0	2	4
	24DBMB21	Basics of Biochemistry	4	0	0	2	4
DSE 2	24DBMB22	Biosafety and Intellectual Property Rights (IPR)	4	0	0	2	4
DSE 3	24DBMB31	Cell culture and Fermentation Technology	3	0	0	2	3
	24DBMB32	Food Fermentation Techniques	3	0	0	2	3
	24DBMB41	Medical virology	4	0	0	2	4
DSE 4	24DBMB42	Antimicrobial Agents and Resistance	4	0	0	2	4
DSE 5	24DBMB51	Environmental Biotechnology	4	0	0	2	4
DSF 2	24DBMB52	Microbiome and Human health	4	0	0	2	4
DSE 6	24DBMB61	Microbiology and Public Health	3	0	0	2	3
DSE 0	24DBMB62	Bioenergetics in Microorganisms	3	0	0	2	3
DSE 7	24DBMB71	Recombinant DNA Technology	4	0	0	2	4
	24DBMB72	Microbial bioremediation	4	0	0	2	4
DSE 8	24DBMB81	Microbial Technology for Human Welfare	4	0	0	2	4
DBEO	24DBMB82	Microbial Nanotechnology	4	0	0	2	4

DISCIPLINE SPECIFIC ELECTIVE COURSES

AECC & LANGUAGES

Category	Code	Course	L	Т	Р	0	С
LANG 1	24LTAM11/ 24LHIN11/ 24LFRE11	Tamil I / Hindi I/ French I	2	0	0	1	2
ENG 1	24LENG11	English I	2	0	0	1	2
LANG 2	24LTAM21/ 24LHIN21/ 24LFRE21	Tamil II / Hindi II / French II	2	0	0	1	2
ENG 2	24LENG21	English II	2	0	0	1	2
LANG 3	24LTAM31/ 24LHIN31/ 24LFRE31	Tamil III / Hindi III / French III	2	0	0	1	2
ENG 3	24LENG31	English III	2	0	0	1	2
AECC 1	24EVS031	Environmental Studies	3	0	0	2	3

MULTIDISCIPLINARY COURSES

Category	Code	Course	L	Т	Р	0	C
MDC 1	24MBMB11	Microbiology of Air and Water	3	0	0	2	3
MDC 2	24MBMB21	Bioinstrumentation	3	0	0	2	3
MDC 3	24MBMB31	Microbial Biotechnology	3	0	0	2	3

MULTIDISCIPLINARY ELECTIVE COURSES

Category	Code	Course	L	Т	Р	0	С
MDE 1	-	Indian Knowledge System	1	1	0	1	2

VALUE ADDED COURSES

Category	Code	Course	L	Τ	Р	0	С
VAC 1	24DVAC11	Universal Human Values	1	0	0	1	1
VAC 2	24DVAC21	Communication Skills	2	0	0	1	2
VAC 3	24DVAC41	Yoga Education / NSS / NCC	0	0	2	1	1
VAC 4	24DVAC51	Spirulina Cultivation	2	0	0	1	2

SKILL ENHANCEMENT COURSES

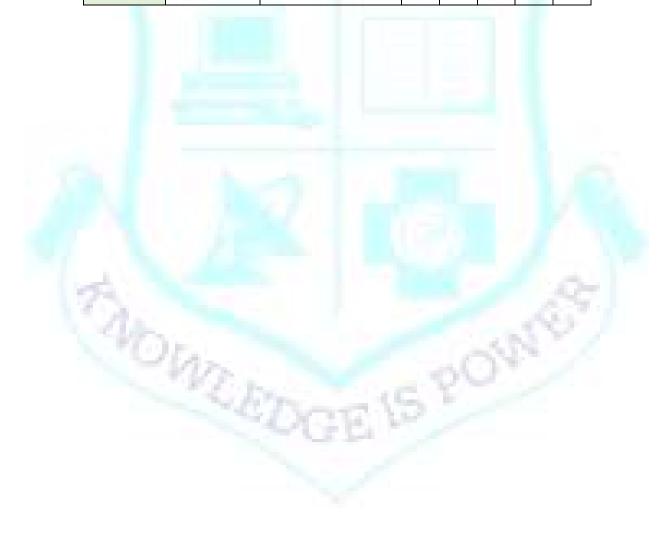
Category	Code	Course	L	Τ	Р	0	С
SEC 1	24SSKU11	Soft Skills I	2	0	0	1	2
SEC 2		Orientation Programme / Industrial Visit	-	-	-	-	-
SEC 3	24SSKU21	Soft Skills II	2	0	0	1	2
SEC 4	24SSKU31	Soft Skills III	2	0	0	1	2
SEC 5	24SBMB41	Industry Oriented Employability skills	1	0	2	1	2
SEC 6	1	In-plant Training/ Industrial Tour/ Summer Term	-	-	-	-	5
SEC 7	24SBMB51	Entrepreneurial Development	2	0	0	1	2
SEC 8	NVII I	Skill Enhancement Training / Student Club Activities/ Institution Innovation Council Activities	5	SC -	2	1	-
SEC 9	24SBMB61	Mini Project	0	0	4	1	2
SEC 10		On Job Training / Apprenticeship / Startup	-	-	-	-	-

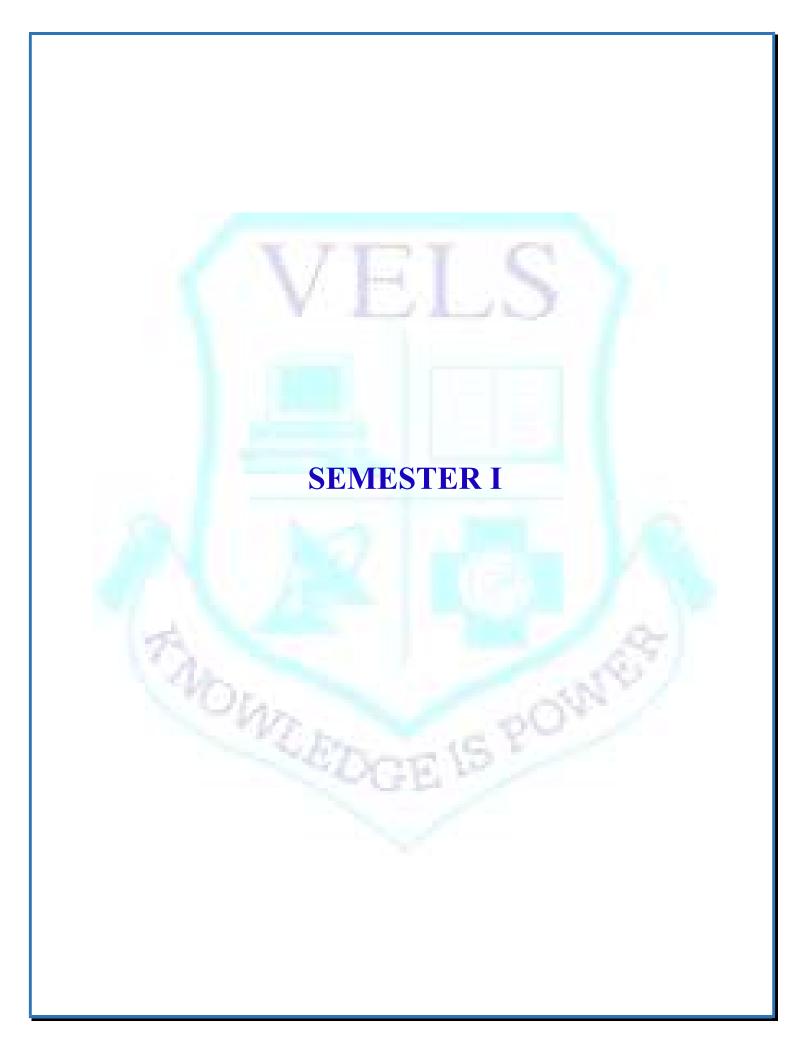
SUMMER INTERNSHIP

Category	Code	Course	L	Τ	Р	0	С
SI 1	24IBMB31	Internship I	0	0	2	1	1
SI 2	24IBMB51	Internship II	0	0	2	1	1

RESEARCH PROJECT

Category	Code	Course	L	Т	Р	0	С
RP 1	24RBMB71	Research Project I	0	0	12	4	6
RP 2	24RBMB81	Research Project II	0	0	12	4	6





24LTAM11 மொழிவரலாறு – சங்க இலக்கியம் – அற இலக்கியம் – மொழித்திறன்

L	Т	Р	0	С
2	0	0	1	2

பருவம்-1, தமிழ்மொழிப்பாடம்-1, பகுதி-1, தகுதிப்புள்ளி: 2, வாரப் பாட நேரம்: 2.

தாள்-1

பாடத்திட்ட நோக்கம்:

மாணவர்களின் இலக்கிய நாட்டத்தை மேம்படுத்துதல், தற்கால தமிழ் மரபுக்கவிதை, புதுக்கவிதை, இலக்கிய வகைமைகளான உரைநடை ஆகியவற்றை அறிமுகப்படுத்துதல், தமிழர்தம் வாழ்வியல் நெறிகளையும் அறியச் பண்பாட்டுச் செழுமைகளையும் இன்றைய தலைமுறையினர் செய்தல், மாணவர்களுக்குத் தமிழைத் தவறின்றி எழுதுவதற்குத் தேவையான பயிற்சி அளித்து அவர்களின் மொழித்திறனை மேம்படுத்துதல், செய்யுளின் நலத்தைப் பாராட்டும் முறைமையை அறியச் செய்து அதன்வழி சிந்தனை வளத்தைப் பெருகச் செய்தல் என்பனவும் மேற்கண்டவழி மாணவர்களை ஆளுமை மிக்கவர்களாக உருவாக்கி, போட்டித்தேர்வுகளுக்குத் தயார் செய்து அவர்களுக்கு வேலைவாய்ப்பை உருவாக்குவதும் இந்தப் பாடத்திட்டத்தின் முக்கிய நோக்கமாகும்.

அலகு - 1: தமிழ் மொழி வரலாறு

08 மணி நேரம்

மொழிக்குடும்பம் - இந்திய மொழிக்குடும்பங்கள் - இந்திய ஆட்சி மொழிகள் - திராவிட மொழிக்குடும்பங்கள் - திராவிட மொழிகளின் வகைகள் -திராவிட மொழிகளின் சிறப்புகள் - திராவிட மொழிகளின் வழங்கிடங்கள் -திராவிட மொழிகளுள் தமிழின் இடம் - தமிழ்மொழியின் சிறப்புகள் - தமிழ் பிறமொழித் தொடர்புகள்.

அலகு -2

புறநானூறு- பாடல் எண்: , 182, 183, - இரண்டு பாடல்கள். குறுந்தொகை- பாடல் எண்: 2, 167, - இரண்டு பாடல்கள் பரிபாடல் - முருகன். வையை - இரண்டு பாடல்கள்

அலகு -3 அற இலக்கியங்கள்

திருக்குறள்- வான்சிறப்பு (அறம்), பெருமை (பொருள்), பிரிவாற்றாமை (இன்பம்),. மூன்று அதிகாரங்கள் முழுமையும் நாலடியார் - இரண்டு பாடல்கள். (2, 3) மூதுரை - இரண்டு பாடல்கள். (2, 8)

07 மணி நேரம்

08 மணி நேரம்

அலகு 4 மொழி

07 மணி நேரம்

பிழை நீக்கி எழுதுதல் - ஒற்றுப்பிழை நீக்கி எழுதுதல் - தொடர்பிழை நீக்கி எழுதுதல் - ஒற்று மிகும் இடங்கள் - ஒற்று மிகா இடங்கள் - பிற மொழிச் சொற்களை நீக்கி எழுதுதல் - பயிற்சிகள்.

மொத்தம்: 30 மணி நேரம்

<u>பார்வை நூல்கள்</u>

தமிழர் நாகரிகமும் பண்பாடும், டாக்டர் அ. தட்சிணாமூர்த்தி, ஐந்திணைப் பதிப்பகம், 2001.

தவறின்றித் தமிழ் எழுதுவோம், மா. நன்னன், ஏகம் பதிப்பகம், 1999.

தவறின்றித் தமிழ் எழுத - மருதூர் அரங்கராசன், ஐந்திணைப் பதிப்பகம், 2003.

தமிழ் இலக்கிய வரலாறு, வரதராசன், மு., புது தில்லி : சாகித்திய அக்காதெமி, 2002.

புதிய தமிழ் இலக்கிய வரலாறு, நீல. பத்மநாபன், சிற்பி பாலசுப்ரமணியம், சாகித்திய அகாடெமி, 2007.

செம்மொழி தமிழின் சிறப்பியல்புகள் - முனைவர் மறைமலை இலக்குவனார்; <u>https://www.youtube.com/watch?v=HHZnmJb4jSY</u>

<u>பாடநூல் தேடலுக்கான இணையம் - https://archive.org/</u>



24LHIN11

HINDI

		L	Т	Р	0	С
		2	0	0	1	2
	वर्ष I – सत्र I (गद्य, पत्र लेखन & व्यावहारिक हिन्दी)	L				
	I Year-Sem I (Prose, Letter Writing & Functional Word	ds)				
Course O	bjectives :					
	understand the rural life style, social responsibilities and	SOC	ial	val	ues	<u>s</u>
	create awareness about the importance of varied culture	1.	1			
	enable the students to develop communication skill in Hin	<u>dı a</u>	nd	to	us	<u>e</u>
AZI	nagi, Azhagi+ fonts					
UNIT I :	पं.श्रीराम शर्मा कृत 'स्मृति' (कहानी)					
	'Smruti' (Kahani) by Pandit Sriram Sharma.			(6hr	S.
UNIT II :	शरद जोशी कृत 'अतिथि तुम कब जावोगे' (व्यंग्य)					
UNII II .	'Athiti tum kab jaaoge' (Vyangy) by Sharad Joshi.				6h	rs.
UNIT III	राहुल सांस्कृतयायन कृत 'अथातो घुमक्कड़ जिज्ञासा' (यात्रा वृत्तान्त)					
	'Atatho Ghumakkad Jigyasa' (Yatra Vruthanth) by			(5hr	'S.
	Rahul Sanskrityayan.					
UNIT IV:	व्यावहारिक हिन्दी- पत्र लेखन में प्रयुक्त वाक्यांश, कौशल विकास					
	– भाव एक भाषा अनेक			(5hr	s.
	Functional Hindi-Phrases used in Letter Writing.					
	Skill development - Bhav Ek Bhasha Anek					
UNIT V :	पत्र लेखन – परिचय व प्रकार, 3 अनोपचारिक पत्र					
	अलगी, अलगी + फ़ॉन्ट का परिचय	ß	2			
	Letter Writing- Intro. & Types & 3 Personal Letters			6	hrs	5.
	Introduction to Azhagi, Azhagi + fonts					
Total: 30	hrs.					
Course O	utcome:					

At the end of this course Students will be able to

- CO1 Gain knowledge about the rural life style
- CO2 Understand social values
- CO3 Understand importance of varied culture
- CO4 Journalise in Functional Hindi
- CO5 Use Azhagi, Azhagi+ fonts

Text Books:

- 1. Pandit Shriram Sharma, Shikaar, (1932) Sahitya Sadan.
- 2. Sharad Joshi, Yatha Sambhav, (2014)Bharatiya Gyanpeet.
- 3. Rahul Sanskrityayan, Ghumakkad Shastra, (1949)Rajkamal Prakashan.

<u>Reference Book:</u>

- 1. Kendriya Hindi Sansthan, (2012) Banking Hindi Patyakram.
- 2. NCERT, Sparsh, Class 9.
- 3. Main Aur Mera Vyakaran, New Saraswati House, New Delhi.
- 4. Govind Ballabh Sharma, (2022) Hindi Vyavaharik Tankan Kala Evam Tankan Abhyas, Neelkanth Publishers Pvt. Ltd.,

<u>Weblinks :</u>

Pandit Sriram Sharma ka kahani: https://www.evidyarthi.in Harishankar parasayi ka Vyangy: http://gadyakosh.org Rahul Sanskrityayan ka yatravruttant: https://www.hindwi.org Prayojanmoolak Hindi:https://hi.mwikipedia.org https://www.azhagi.com/hnd/helphtml/Introduction.html

MONDET

FRENCH PAPER I

L	Т	Р	0	С
2	0	0	1	2

24LFRE11

Course Objectives	
The lessons are being chosen:	
1) to greet, to express excuse and to introduce oneself	
2) to introduce another person	
3) to express his/her ideas, opinions and weekend projects	
4) to request someone to do something, polite manners	
5) to accept, refuse, enquire and indicate the time and date	
6) to express himself / herself in positive and negative manner	
UNITS	
1) Salut	
les nombres, Les jours de la semaine et du mois, La nationalité	4 hours
2) Enchanté	
Les verbes Etre, Avoir, Aller, Regular ER verbes, Present tense.	6 hours
3) J'Adore	
La negation, l'adjectif possessif, le futur proche	4 hours

4) Tu veux bien

Les articles de finis/indéfinis, Les pronoms après une préposition (avec lui, chez moi), Le passé composé 7 hours

5) On se voit quand

Les pronoms compléments directs me, te, nous, vous, L'interrogation avec est-ce que, L'heure et la date. 5 hours

6) Bonne idée

Les articles partitifs, Le masculin et le féminin des adjectifs, Les pronoms compléments directs le, la, les, La négation : ne... pas de. 4 hours Total no. of hours - 30 hours

Course Outcome :

- 1) The students would be able to greet, to excuse and to introduce himself
- 2) The students would be able to introduce someone
- 3) The students would be able to express his ideas, opinions and weekend projects
- 4) The students would be able to ask someone to do something, polite manner
- 5) The students would be able to accept, refuse enquire and indicate the time and date
- 6) The students would be able to express himself in positive and negative manner

Text / Reference Book:

Prescribed book: LATITUDES 1 (A1/A2) MÉTHODE DE FRANÇAIS - Régine Mérieux and Yves Loiseau

Reference book: SAISON A1 - MÉTHODE DE FRANÇAIS - Marie-Noëlle Cocton, Élodie Heu, Catherine Houssa, Émilie Kasazian

24LENG11

ENGLISH - I

COURSE OBJECTIVES:

- To enable students to develop their communication skills effectively.
- To make students familiar with usage skills in the English Language.
- To enrich their vocabulary in English.
- To develop communicative competence.

Unit I- Prose	Credit Hours
• Dangers of drug abuse - Hardin B.Jones	06
• Tight corners - E.V.Lucas	
Unit II -Poetry	06
 Ecology - A.K.Ramanujan 	
• The owl and the chimpanzee - Jo Camacho	
Unit III - Short story	06
• The Dear Departed - Stanley Houghton	
• The Fool's Paradise- Isaac Bashevis Singer	
Unit IV -Grammar	06
• Parts of speech, Articles	
Unit V -Grammar	06
• One-word substitution, prefix, suffix, synonym, antonym.	
	Total 30 hours

Course Outcomes:

At the end of this course, the students would have learnt to

CO1	Understand the characteristic features of the language used in the text.			
CO2	Strengthen their knowledge of basic grammar			
CO3	Improve narrative skills after studying diverse prose and play.			
CO4	Understand to classify parts of speech and articles.			
CO5	Develop critical writing skills in the textual content of the syllabus.			

<u>References:</u>

- English for Communication Enrichment: by Jeya Santhi June 2015.
- Dr. M. Narayana Rao and Dr. B. G.Barki–Anu's Current English for Communication (AnuChitra). June 2012.
- Dr. Ananthan, R. Effective Communication. Ed. Chennai: Anu Chithra Pub.2010.

L	Т	Р	0	С
2	0	0	1	2

24CBMB11 BASIC TECHNIQUES IN MICROBIOLOGY COURSE OBJECTIVE:

The candidate will gain hands-on knowledge and acquire adequate skill

required to sterilize media and to prepare, inoculate observe and distinguish the growth patterns in different media.

UNIT-1 INTRODUCTION TO MODERN MICROSCOPY

Bright field microscopy: Electromagnetic spectrum of light, Structure, working of and ray diagram of a compound light microscope, Types, ray diagram and functions of – condensers (Abbe and cardioid) eyepieces and objectives, Concept of aberrations in lenses - spherical, chromatic, comma and astigmatism, Principle, working and ray diagram of - Phase contrast microscope, Fluorescence Microscopy, Electron Microscopy – TEM, SEM.

UNIT -11 STAINING TECHNIQUES

Definition of Stain; Types of stains (Basic and Acidic), Properties and role of Fixatives, Mordants, Decolourisers and Accentuators, Monochrome staining and Negative (Relief) staining, Differential staining - Gram staining and Acid-fast staining, Special staining- Capsule, Cell wall, Spore, Flagella, Lipid granules, metachromatic granules.

UNIT-111 STERILIZATION AND DISINFECTION

Sterilization: Physical Agents - Heat, Radiation, Filtration, Checking of efficiency of sterilization (Dry and Moist) – Biological and Chemical Indicators, Disinfection: Chemical agents and their mode of action - Aldehydes, Halogens, Quaternary ammonium compounds, Phenol and phenolic compounds, Characteristics of an ideal disinfectant, Heavy metals, Alcohol, Dyes, Detergents and Ethylene oxide, Checking of efficiency of disinfectant - Phenol Coefficient (Rideal–Walker method).

UNIT-1V CULTIVATION OF MICROORGANISMS

Nutritional requirements and nutritional classification, Design and preparation of media: Common ingredients of media and types of media, Methods for cultivating photosynthetic, extremophilic and chemo-lithotrophic bacteria, anaerobic bacteria, algae, fungi, actinomycetes and viruses, Concept of Enrichment, Pure Culture, Isolation of culture by streak plate, pour plate, spread plate, Maintenance of bacterial and fungal cultures using different techniques, Culture collection centres and their role, Requirements and guidelines of National Biodiversity Authority for culture collection centres

UNIT-V BACTERIAL GROWTH

Kinetics of bacterial growth (Exponential growth model), Growth curve and Generation time, Diauxic growth, Measurement of bacterial growth- Methods of enumeration: Microscopic methods

L	Т	Р	0	С
3	0	0	2	3

9

9

9

9

9

(Direct microscopic count, counting cells using improved Neubauer, Petroff-Hausser's chamber), Plate counts (Total viable count), Turbidometric methods (including Nephelometry), Estimation of biomass (Dry mass, Packed cell volume), Chemical methods (Cell carbon and nitrogen estimation), Factors affecting bacterial growth {pH, Temperature, Solute Concentration (Salt and Sugar)} and Heavy metals

Total: 45 Hours

Course Outcome:

At the end of this course students will be able to:

- **CO1:** Develop skills to observe microbes using microscopes.
- CO2: Competently prepare and cultivate bacteria in different types of media.
- CO3: Judge the accuracy of sterilization
- CO4: Prepare media and grow fungi and algae in the lab
- CO5: Evaluate the sensitivity and resistance of bacteria to antimicrobials
- **CO6:** Explain the principles behind common microbiological techniques (e.g., staining methods, culturing techniques).

CO7: Summarize the differences between various types of microbial cultures.

<u>Reference Books:</u>

- T. Michael, M. Madigan, John. P. Martinko, David. Stahl, and A. David Clark (2010), Brock Biology of Microorganisms" 13th Edition. This textbook offers a detailed introduction to microbiology, including fundamental techniques and applications. It covers various microbiological methods and the principles behind them.
- J. Gerard. Tortora, R. Berdell. Funke, and L. Christine. Case (2013). Microbiology: An Introduction" 11th Edition. This book provides a thorough introduction to microbiological techniques and concepts, including practical laboratory methods and their applications.
- C. Karen. Carroll, C. Janice. Love, and A. Michael. P. faller (2023). Manual of Clinical Microbiology This manual is a comprehensive reference for clinical microbiology techniques, including culture methods, staining, and identification procedures.
- Albert Balows, J. William. J. Hausler, Karl Hermann, and L. Kenneth. Herrmann (2006). Practical Handbook of Microbiology" 3rd Edition. A practical guide that covers essential microbiological techniques and protocols. It is useful for both beginners and experienced practitioners.
- J.G. Holt, N.R. Krieg, P.H.A. Sneath, J. Staley, and S.T. Williams (2004). Microbiological Methods" 8th Edition. Description: This book provides detailed methods for microbiological laboratory techniques, including culturing, staining, and biochemical testing.

24CBMB12 GENERAL MICROBIOLOGY

COURSE OBJECTIVE:

The candidate will gain knowledge about the structure of bacteria, fungi, algae, protozoa and viruses along with the basic principles of microscopy. Control of microbial growth by physical and chemical methods plus the use of antibiotics and their efficacy testing are emphasized. Cultivation of microbes and space microbiology is discussed.

UNIT-I INTRODUCTION

Historical perspective and scope of microbiology. Description of various groups of microorganisms with typical example. Cell cycle and reproduction of bacteria. Bacterial cell structure and components. Microbial classification. Tools and techniques in microbiology

UNIT-II MICROSCOPY

Microscopy-principles of microscopy-bright-field microscopy-Simple microscope, Compound microscope, PCM, FM, DFM,TEM,SEM and STEM-description, principle and use. Scanning Probe Microscopy (SPM), Applications of Microscopy, Preparing Samples for Microscopy, Modern Advances -Super-Resolution Microscopy, Cryo-Electron Microscopy (Cryo-EM)).

UNIT-III STERILIZATION

Sterilization– Principle, use and quality control of: High temperature -Dry Heat- Hot air oven, incineration, moist heat- Tyndallization, Pasteurization, inspissation, moist heat under pressure; low temperature–preservation; filtration-membrane filters, depth filters; centrifugation; radiation. Disinfection-Mode of action and Evaluation – RW test, KS Test, Use-Dilution method. Sterilization Validation (Biological Indicators, Chemical Indicators, Mechanical Indicators)

UNIT – IV ANTIBIOTICS

Antibiotics–Classification, Mode of Action, mechanism of resistance, Evaluation–Disc Diffusion; MIC– Broth dilution, agar dilution; MBC; E-test with Quality control for each method. Development of New Antibiotics -Novel Antibiotics, Teixobactin, Murepavadin, Rapid Diagnostic Techniques -Point-of-Care Testing, Genomic and Proteomic Approaches, Microbiome-Based Therapies.

UNIT-V MICROBIAL GROWTH

Growth requirements of Bacteria. Microbial culture media and pure culture techniques.

L	Т	Р	0	С
4	0	0	2	4

12

12

12

12

12

Anaerobic cultivation methods. Preservation of cultures. Quantitation of microbial growth and bacterial growth curve in batch culture. Microbial Growth in Extreme Environments - Thermophiles, Space Microbiology.

Total: 60 Hours

Course Outcome:

At the end of this course students will be able to:

- **CO1**: Demonstrate knowledge of fundamental microbiological principles.
- CO2: Evaluate the outcomes of various antibiotic sensitivity tests.
- CO3: Decide the optimum media for growth of microbes.

CO4: Assess the outcome of sterilization and disinfection protocol.

CO5: Judge the suitability of microscopes to microbial cytology.

CO6: Choose appropriate methods for cultivation of microorganisms.

Text Books:

1. J. Michael, E.C.S.Chan, Noel R.Krieg (2006), Microbiology, Tata-McGrawHill.Ed.5.

Reference Books:

- R. Ananthanarayanan & C.K.Jeyaram Paniker (2005), Textbook of Microbiology, Ed.7, 2. Orient Longman.
- T. Michael T. Madigan, M. John Martinko (2006), Brock's Biology of Microorganisms, Ed.11 Pearson-PrenticeHall.
- 3. M. Ronald Atlas (1997), Principles of Microbiology, Ed. 2. WCB Publishers.
- Y. Roger, John Stanier, L. Ingraham, L. Mark Page Wheelis, Page R. Painter (2004), General Microbiology, Ed. 5. MacMillan Press.
- Topley & Wilson's (2002), Principles of Bacteriology, Virology&Immunology, Ed. 9, Edward Arnold.
- M. Lansing Prescott, P. John Harley, Donald A. Klein (2005), Microbiology, McGrawHill.Ed.6.
 <u>Web Sources:</u>
 - 1. https://www.britannica.com/science/microbiology
 - 2. https://www.slideshare.net/slideshow/lect-1-introduction/34565343
 - 3. https://www.slideshare.net/slideshow/introduction-to-microbiology-67925181/67925181

24MBMB11 MICROBIOLOGY OF AIR AND WATER

COURSE OBJECTIVE:

The candidates will gain comprehension of microbial interactions with the environment, specifically in the realms of air and water, as well as their correlation with diseases. The students will also understand the significance of microorganisms as indicators of pollution. The purpose of this syllabus is to give students a thorough comprehension of the microbial ecology of air and water, with a focus on both theoretical understanding and practical implementation in the fields of environmental and public health.

UNIT- I INTRODUCTION TO MICROBIOLOGY OF AIR AND WATER

Bio aerosols, Air borne microorganisms (bacteria, Viruses, fungi) and their impact on human health and environment, significance in food and pharma industries and operation theatres, allergens. Brief account of air borne water borne transmission of harmful microbes.

UNIT-II AIR SAMPLE COLLECTION AND ANALYSIS NO OF HOURS 9

Bio aerosol sampling, air samplers, methods of analysis, CFU, culture media for bacteria and Fungi, Identification characteristics. Microbial diversity in air and water. Method of air sampling and types of air samplers; Impaction on solids, Impingement in liquid, Sedimentation and Centrifugation. Air sanitation: Physical and chemical methods.

UNIT-III WATER MICROBIOLOGY

Types of water: surface, ground, stored, distilled, mineral and demineralized water. Recommended Bacteriological standards of Water Quality, Tamil Nadu pollution control board (TNPCB). Main Functions of TNPCB. Water quality standards for best designated usages, Central pollution control board, (CPCB), Main Functions of CPCB.

UNIT – IV WATER TREATMENT AND PURIFICATION

Water purification methods. Water borne Infections; Indicators of faecal pollution, Escherichia coli, Bifidobacterium, Streptococcus faecalis, Clostridium perfringens. New indicators: Campylobacter and Pseudomonas. Environmental Factors Affecting Microbial Populations; BIS and WHO.

UNIT-V **APPLIED MICROBIOLOGY OF AIR AND WATER**

Microbial bioremediation of air and water pollutants, Role of microbes in sustainable environmental practices. Control Measures; Precipitation, chemical disinfection, filtration, high temperature and UV light.

Total: 45 Hours

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Course Outcome:

At the end of this course students will be able to:

CO1: Understand the fundamental principles of microbiology as they apply to air and water environments.

CO2: Identify and classify the types of microbes present in air and water and their ecological roles.

CO3: Evaluate the impact of environmental factors on microbial populations in air and water.

CO4: Analyze the significance of microbes in the context of public health and environmental sustainability.

CO5: Apply knowledge of microbiological principles to solve real-world problems related to air and water quality.

CO6: Perform techniques for the isolation and identification of microorganisms from air and water samples.

Text Books:

1. A Martin (1961), Introduction to Soil Microbiology, John Wiley& Sons, New York and London Publication.

- 2. N. S Subba Rao (1977), Soil Microbiology, 4th Ed., Oxford & IBH Publishing Co. Pvt. Ltd.
- 3. R.C Dubey and D. K Maheswari, (2023), Textbook of Microbiology, S. Chand & Co.
- A Martin (1977), An Introduction to Soil Microbiology. 2nd edition. John Wiley & Sons Inc. New York & London.
- H.C Dube and K.S Bilgrami (1976), Text book of modern pathology. Vikas publishing house. New Delhi.
- 6. G Rangaswami (1979), Recent advances in biological nitrogen fixation. Oxford and IBH. New Delhi.
- 7. Daniel Lim (2003), Microbiology, 2nd Edition; McGraw-Hill Publication.
- 8. J.L Ingraham and C.A Ingraham (2004), Introduction to Microbiology. 3nd Edition. Thomson Brooks / Cole.
- M.T Madigan, J.M Martinko (2006), Brock's Biology of Microorganisms. 11th Edition. Pearson Education Inc.

Reference Books:

- N da Silva, MH Taniwaki, VC Junqueira N Silveira, MS Nascimento, RAR Gomes (2012) Microbiological Examination Methods of Food and WaterA Laboratory Manual, CRC Press.
- 2. RM Atlas and R Bartha (2000), Microbial Ecology: Fundamentals & Applications. 4th edition. Benjamin/Cummings Science Publishing, USA

- 3. RM Maier, IL Pepper and CP Gerba (2009), Environmental Microbiology. 2nd edition, Academic Press.
- 4. C.J. Hurst, R.L. Crawford, J.L. Garland, D.A. Lipson (2007), Manual of Environmental Microbiology, 3rd edition, ASM press.

Web Sources:

1. Home - Books - NCBI (nih.gov) 2. Food Microbiology Notes - Microbe Notes NOWLEI 520

24DBMB11 MICROBIAL DIVERSITY

COURSE OBJECTIVE:

This course provides an in-depth exploration of microbial diversity, encompassing bacteria, archaea, fungi, viruses, and protists. Emphasis will be placed on understanding the ecological roles, evolutionary relationships, and applications of microorganisms in diverse environments.

UNIT-I INTRODUCTION TO TAXONOMY

Introduction to taxonomy; Taxonomical hierarchy, Binomial Nomenclature; concept of species; Major characteristics used in classification; five kingdom, six kingdom and eight kingdom concepts.

UNIT-II CLASSIFICATION OF BACTERIA

Morphological Basic classification of Bacteria and Basis for Classification of bacteria according to Bergey's Manual of Systematic Bacteriology.

UNIT-III CLASSIFICATION OF ALGAE

Classification of Algae, Structure of algal cell with example; Life-cycle patterns of Algae. Reproduction in algae. Structure of Fungi - unicellular and multicellular forms. Industrial impotent Algae

UNIT – IV CLASSIFICATION OF VIRUSES

Classification of animal and Plant viruses, Structure of Virus – Adenovirus, Retrovirus, TMV and T- phage. Replication of Viruses – RNA Viruses ssRNA, ds RNA and DNA Viruses ssDNA, dsDNA.

UNIT-V CLASSIFICATION OF FUNGI

General Characteristics of fungi; Range of thallus organization and Reproduction. Classification according to Alexopolous, Mims and Black Well. General characteristics of Fungi.

Total: 60 Hours

Course Outcome:

At the end of this course students will be able to:

- **CO1:** Define microbial diversity and different types of microorganisms.
- CO2: Recall key characteristics of bacteria, archaea, fungi, viruses, and protists.
- **CO3:** List methods used to study microbial diversity, such as sequencing, microscopy, and culture techniques.

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CO4: Explain the ecological roles of microorganisms in nutrient cycling, decomposition, and symbiosis.

CO5: Summarize the evolutionary processes that have shaped microbial diversity.

CO6: Interpret scientific literature on microbial diversity and ecosystems.

CO7: Apply knowledge of microbial diversity to analyze case studies of microbial communities in specific environments (e.g., soil, oceans, human microbiome).

CO8: Evaluate the effectiveness of different techniques for studying microbial diversity.

CO9: Assess the implications of microbial diversity research for human health, agriculture, and environmental sustainability.

<u>Text Books:</u>

1. G.J Tortora, B.R. Funke and C.L. Case (2008). Microbiology: An Introduction. 9th edition. Pearson Education

2. M.T. Madigan, J.M. Martinko P.V Dunlap D.P and Clark (2014). Brock Biology of Microorganisms. 14th edition. Pearson International Edition

<u>Reference Books:</u>

J.Cappucino and N.Sherman (2010). Microbiology: A Laboratory Manual. 9th edition.
 Pearson Education Limited

2. J.M.Wiley, L.M.Sherwood C.J.and Woolverton (2013) Prescott's Microbiology. 9th Edition. McGraw Hill International.

3. R.M. Atlas (1997). Principles of Microbiology. 2nd edition. WM.T.Brown Publishers.

4. M.J. Pelczar, E.C.S Chan and Krieg N.R. (1993). Microbiology. 5th edition. McGraw Hill Book Company.

5. R.Y. Stanier , J.L.Ingraham, M.L. Wheelis, and P.R.Painter (2005). General Microbiology. 5th edition. McMillan.

24DBMB12

MICROBIAL PHYSIOLOGY AND METABOLISM

COURSE OBJECTIVE:

This course provides an in-depth exploration of microbial cell structure, function, and physiology. Explore the diverse metabolic pathways utilized by microorganisms for growth and energy production. Analyze the regulatory mechanisms governing microbial metabolism under different environmental conditions.

UNIT- I MICROBIAL CELL STRUCTURE AND FUNCTION

Overview of microbial cell structure: Bacteria, Archaea, and Eukaryotes. Functions of cell membranes, cell walls, and cellular organelles. Growth phases, cell division, and nutrient transport mechanisms. Energy production: ATP synthesis and electron transport chain.

UNIT-II MICROBIAL METABOLISM – CATABOLISM

Glycolysis, TCA cycle, and fermentation pathways, Anaerobic and aerobic respiration. Bioenergetics: Oxidative phosphorylation and chemiosmosis. Metabolism of carbohydrates, lipids, and proteins.

UNIT-III MICROBIAL METABOLISM – ANABOLIC PATHWAYS

Biosynthesis of amino acids, nucleotides, and fatty acids. Assimilation of nitrogen, sulfur, and phosphorus Anaplerotic reactions and glyoxylate cycle Regulation of anabolic processes

UNIT – IV ENVIRONMENTAL INFLUENCES ON MICROBIAL PHYSIOLOGY

Influence of temperature, pH, osmotic pressure, and oxygen levels on growth. Microbial stress responses: Heat shock, oxidative stress, and nutrient starvation. Adaptation to extreme environments: Thermophiles, halophiles, acidophiles. Quorum sensing and microbial communication.

UNIT– V APPLICATIONS OF MICROBIAL PHYSIOLOGY AND METABOLISM 12

Industrial fermentation and bioprocessing. Microbial biotechnology: Production of enzymes, antibiotics, and biofuels. Environmental applications: Bioremediation and biodegradation Use of microbial physiology in synthetic biology and metabolic engineering.

Total: 60 Hours

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Course Outcome:

At the end of this course students will be able to:

CO1: Students will demonstrate a thorough understanding of the physiological processes governing microbial growth, metabolism, and adaptation.

CO2: Students will be able to recognize the diversity of microorganisms and their adaptations to various environmental conditions.

CO3: Students will be able to classify microorganisms based on their metabolic characteristics, including chemotrophs, phototrophs, autotrophs, and heterotrophs, and their contribution to microbial diversity.

CO4: Students will analyse microbial interactions and community dynamics, including mutualism, commensalism, parasitism, and competition in terms of their ecological and medical significance.

CO5: Students will be able to appraise the role of biofilms and quorum sensing in microbial communities, as well as the physiological adaptations of extremophiles to harsh environments.

CO6: Students will be able to apply knowledge of microbial physiology to real-world challenges in biotechnology, medicine, environment and environmental remediation.

CO7: Students will be able to solve practical problems in various fields such as industry, biotechnology, medicine, and environment.

Text Books:

- Jr, M.J, Pelczar, N.R. Krieg (1986), Microbiology, 5th edn, McGraw-Hill Book Company, NY.
- J.L Ingraham, and C.L. Ingraham, (2000), Introduction to Microbiology, 2nd edn, Brooks/Cole, Singapore.
- J.G. Black (2002), Microbiology: Principles and Explorations, 5th edn, John Wiley and Sons, Inc. NY.

Reference Books:

- M.T. Madigan and J.M. Martinko (2014), Brock Biology of Microorganisms, 14th edition. Prentice Hall International Inc.
- 2. A.G. Moat and J.W. Foster (2002), Microbial Physiology, 4th edition, John Wiley & Sons.
- 3. S.R..Reddy and S.M.Reddy (2005), Microbial Physiology, Scientific Publishers India.
- 4. G.Gottschalk (1986), Bacterial Metabolism, 2nd edition, Springer Verlag.

- 5. R.Y. Stanier, J.I. Ingrahm, M.L.Wheelis and P.R. Painter (1987), General Microbiology, 5th edition, McMillan Press.
- 6. J.M. Willey, L.M. Sherwood and C.J Woolverton (2013), Precott's Microbiology, 9th edition, McGraw Hill Higher Education.

15 POV

Web Sources:

- 1. https://spot.colorado.edu/~schmidts/Teaching/EPOB3400/microPhys.html
- 2. https://dl.icdst.org/pdfs/files3/0bc9d88695de86f1fbad48fe3fccffc9.pdf

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3. https://nsdl.niscpr.res.in/bitstream/123456789/803/1/CarbonMetabolism.pdf

24PBMB11

PRACTICAL - I

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0	0	2	1	1

PRACTICAL IN BASIC MICROBIOLOGICAL TECHNIQUES

COURSE OBJECTIVE:

The candidate will gain hands-on knowledge and acquire adequate skill required to sterilize media and to prepare, inoculate observe and distinguish the growth patterns in different media.

- 1. Cleaning and Sterilization of Glassware.
- 2. To study the principle and applications of important instruments (biological safety cabinets, autoclave, incubator, BOD incubator, hot air oven, light microscope, pH meter) used in the microbiology laboratory
- 3. Preparation of culture media for bacterial cultivation
- 4. Preparation and growth of Bacteria in Basal Media– Nutrient Broth, Peptone Water, Nutrient Agar.
- 5. Preparation and growth of Bacteria in MacConkey Agar and Cetrimide Agar.
- 4. Preparation and growth of Bacteria in Carbohydrate Fermentation Media.
- 6. Simple staining– positive and negative staining.
- 7. Gram staining of Bacteria (Study of different shapes of bacteria).
- 8. Capsule staining.
- 9. Spore staining.
- 10. Cultivation of fungi in SDA (Study of Rhizopus, Penicillium, Aspergillus) and LPCB.
- 12. Antibiotic sensitivity test KirbyBauer Method.
- 13. Study of the following protozoans using permanent mounts/photographs: Amoeba, Entamoeba, Paramecium and Plasmodium

Total: 45 Hours

Course Outcome:

At the end of this course students will be able to:

CO1: Develop skills to observe microbes using microscopes.

CO2: Competently prepare and cultivate bacteria in different types of media.

CO3: Judge the accuracy of sterilization

CO4: Prepare media and grow fungi and algae in the lab

CO5: Evaluate the sensitivity and resistance of bacteria to antimicrobials

CO6: Explain the principles behind common microbiological techniques (e.g., staining methods, culturing techniques).

CO7: Summarize the differences between various types of microbial cultures.

CO8: Perform basic microbiological techniques accurately and safely in a laboratory setting.

CO9: Demonstrate proficiency in using microscopes and other equipment for microbiological analysis.

24DVAC11

UNIVERSAL HUMAN VALUES

COURSE OBJECTIVE:	COBJECTIVE:
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The candidates will be able to appreciate the complementarity between the values and skills for sustained happiness and prosperity. To influence the students to approach the life and profession with a holistic perspective towards a value-based living in a natural way. To highlight plausible implications of holistic understanding of ethical human conduct.

INTRODUCTION TO VALUE EDUCATION UNIT-I

Living a fulfilling life. Value education. Skill education. Complementarity of Values and Skills. Development of a holistic perspective. Right understanding, relationship and physical facility. Understanding the happiness and prosperity.

UNIT-II HARMONY AT MULTIPLE LEVELS

Human being as co-existence of the self and the human body. Understanding harmony in the self. Harmony in the family and understanding values in human-human relationships. Harmony in the society and understanding universal human order. Harmony in nature and understanding the interconnectedness, self-regulation and mutual fulfillment. Harmony in existence and understanding co-existence at various levels.

UNIT-III IMPLICATIONS OF THE RIGHT UNDERSTANDING

Ethical human conduct. Implications of value-based living. Right understanding of professional ethics. Humanistic education. Holistic technologies, production systems and management models. Strategies for transition towards value-based life and profession.

Total: 15 Hours

Course Outcome:

At the end of the course learners will be able to:

CO1: Develop qualities like responsibility and the ability to handle problems with sustainable solutions.

CO2: Appraise human values and the harmony at various levels.

CO3: Perceive a better critical ability.

CO4: Develop qualities pertaining to value-based living.

CO5: Apply what they have learnt to their own self in real life settings.

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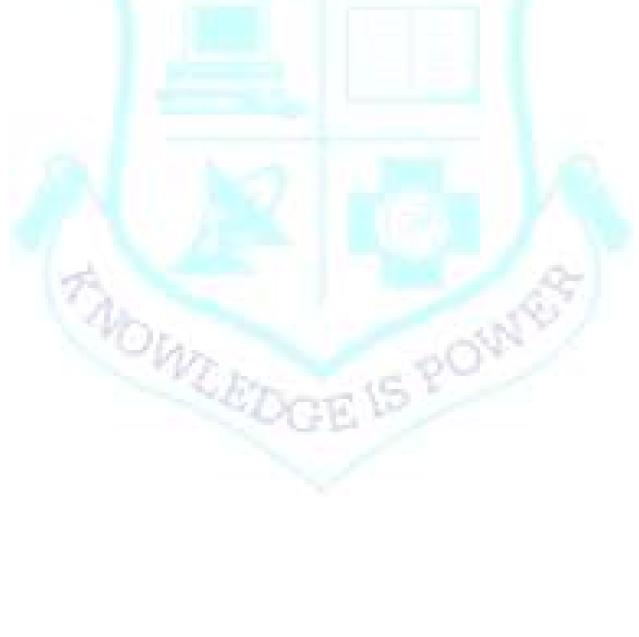
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Text Books:

 R.R. Gaur, R. Asthana, G.P. Bagaria. (2023), A Foundation Course in Human Values and Professional Ethics. 3rd Revised Edition. Excel Books, New Delhi.

Reference Books:

- 1. A. Nagaraj, Jeevan Vidya Prakashan, Amar Kantak. Jeevan Vidya: Ek Parichaya . (1999),
- 2. Rakesh Gupta. (2008), Jeevan Vidya: An Introduction (Introductory Book to Madhyasth Darshan-Coexistentialism). English Version.
- 3. A. N. Tripathi. (2004), Human Values. First Edition. New Age International Publishers, New Delhi.



24SSKU11

SOFT SKILLS -I

COURSE OBJECTIVES:

- To learn and apply basic etiquette for personal and professional interactions.
- To develop effective stress management techniques for maintaining mental and emotionalwell-being.
- To enhance self-awareness for personal growth and informed decision-making.
- To gain an overview of essential 21st-century skills necessary for success in a rapidly changing world.
- To foster creativity and critical thinking skills for innovative problem-solving and adaptability.

UNIT I INTRODUCTION TO SOFT SKILLS

- Soft Skills vs Hard Skills
- 15 important Soft Skills
- Communication Skills, Time Management, Leadership Skills

UNIT II - OVERVIEW OF 21st CENTURY SKILLS.

- Lateral Thinking Left Brain/Right Brain Functionality
- Problem solving skills

UNIT III - SELF AWARENESS

- Human Values
- Mindfulness
- SWOT Analysis
- PDCA Approach

UNIT IV - CREATIVITY/CRITICAL THINKING

- Six Thinking Traits
- Creative writing exercises
- Open mindedness

UNIT V - PERSONAL HYGIENE AND STRESS MANAGEMENT

- Basic Etiquettes
- Health and Personal Grooming
- Stress-meaning and nature, Eustress, Distress
- Stress management strategies

Total Hours30

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Credit Hours

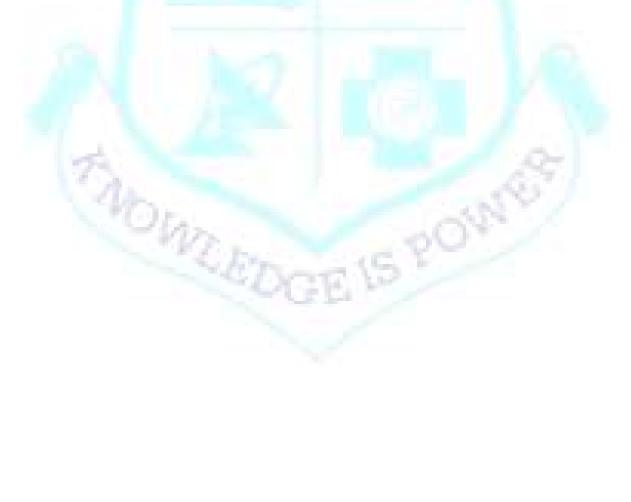
COURSE OUTCOMES:

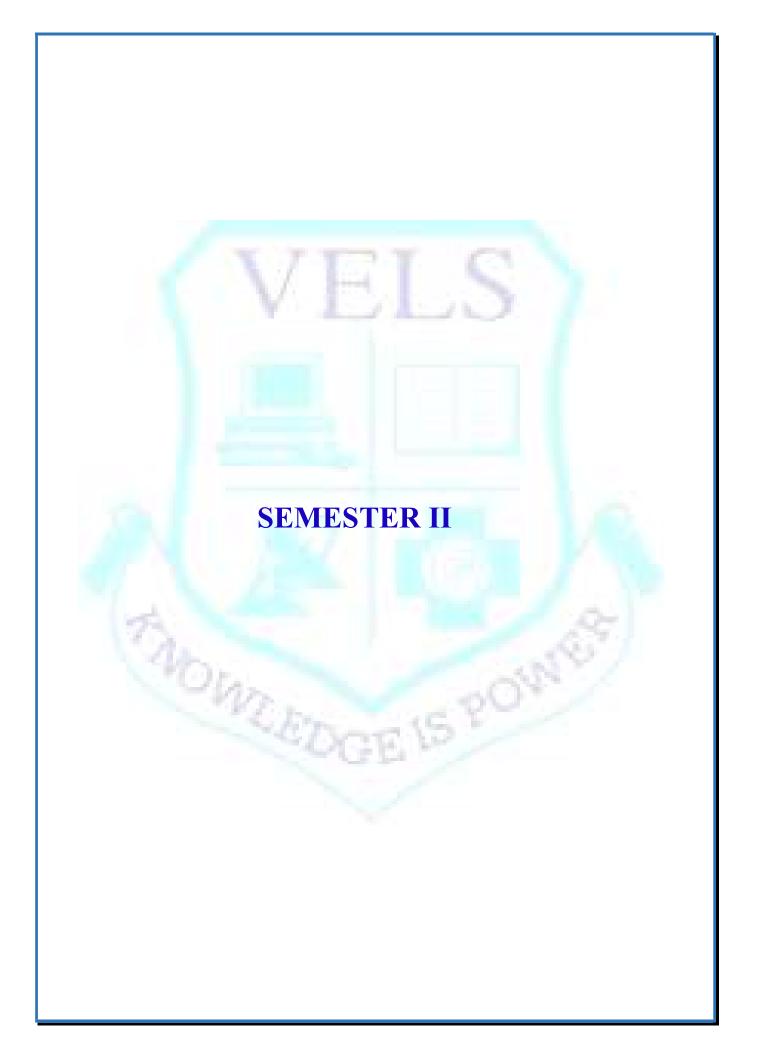
At the end of this course, the students would have learnt to

CO1	demonstrate basic etiquette in various personal and professional settings.
CO2	effectively manage stress using learned techniques.
CO3	show increased self-awareness and make informed decisions.
CO4	understand and articulate key 21st-century skills.
CO5	apply creativity and critical thinking to solve problems innovatively.

<u>References:</u>

- K. Alex (2014). Soft Skills (1st edition) S Chand & Company
- Taylor. (2005) Grant English Conversation Practice. Tata McGraw Hill Education Pvt. Ltd
- Tiko, Champa& Jaya Sasikumar. (1979) Writing with a purpose. OUP New Delhi
- Nelson-Jones, R. (1992). Life skills, a handbook, Trowbridge, Wilts: Detesios Ltd.
- Tuhovsky, Ian (2019). Communication Skills Training (2nd edition) Rupa PublicationIndia.





24LTAM21 பருவம்-2, தமிழ்மொழிப்பாடம்-2, பகுதி-1,

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தகுதிப்புள்ளி: 2, வாரப் பாட நேரம்: 2. தாள்-2

காப்பியம், பக்தி இலக்கியம், கலைகள், நாகரிகம்-பண்பாடு பாடத்திட்ட நோக்கம்:

மாணவர்களின் இலக்கிய நாட்டத்தை மேம்படுத்துதல், அற இலக்கியங்கள், சிற்றிலக்கியம், சிறுகதை ஆகியவற்றை அறிமுகப்படுத்துதல், தற்காலப் பேச்சுத் தமிழ் எழுத்துத்தமிழ் ஆகியவற்றின் வளர்நிலைகளை மாணவர்களை அறியச் செய்தல், அதன்வழி சிந்தனை வளத்தைப் பெருகச் செய்தல் என்பனவும் மேற்கண்டவழி மாணவர்களை ஆளுமை மிக்கவர்களாக உருவாக்கி, போட்டித்தேர்வுகளுக்குத் தயார் செய்து அவர்களின் மொழித் மேம்படுத்த அவர்களுக்குக் கடிதம் எழுதும் கலையைக் திறனை கற்றுக்கொடுத்தல், அணி இலக்கணத்தை அறியச் செய்தல் என்பன இந்தப் பாடத்திட்டத்தின் முக்கிய நோக்கமாகும்.

அலகு 1 காப்பியங்கள்

08மணி நேரம்

சிலப்பதிகாரம்- கனாத்திறம் உரைத்தக் காதை முழுவதும். மணிமேகலை- மலர்வனம் புக்க காதை முழுவதும். கம்பராமாயணம் - குகப் படலம் (தேர்ந்தெடுக்கப்பட்ட ஒன்பது பாடல்கள்) அலகு 2: பக்தி இலக்கியம் 08 மணி நேரம்

- 1. மாணிக்கவாசகர் திருவாசகம் மூன்று பாடல்கள்
 - 🗸 புல்லாகி பூடாகி (சிவபுராணம்)
 - 🗸 எல்லாப் பிறப்பும் (சிவபுராணம்)
 - 🗸 உற்றாரை யான் வேண்டேன் (திருப்புலம்பல்)
- 2. ஆண்டாள் திருப்பாவை மூன்று பாடல்கள் (1, 3, 4)
 - 🗸 மார்கழித் திங்கள் ... (பாசுரம் 1)
 - 🗸 ஓங்கி உலகளந்த... (பாசுரம் 3)
 - 🗸 ஆழிமழைக் கண்ணா... (பாசுரம் 4)

- 3. வீரமாமுனிவர் தேம்பாவணி வளன் செனித்தப் படலம்
- 4. சீறாப்புராணம்- மானுக்கு பிணை நின்ற படலம்

அலகு 3 கலைகள்

07 மணி நேரம்

சிற்பம் - ஓவியம் - இசை - கூத்து - ஒப்பனை - ஆடை அணிகலன்கள்.

அலகு 4 நாகரிகம், பண்பாடு

07மணி நேரம்

சொற்பொருள் விளக்கம் - பண்டைத் தமிழர் வாழ்வியல் - அகம் - களவு - கற்பு -குடும்பம் - விருந்தோம்பல் - உறவு முறைகள் - சடங்குகள் - நம்பிக்கைகள் -பொழுதுபோக்கு - புறம் - போர் முறைகள் - நடுகல் வழிபாடு - கொடைப்பண்பு.

மொத்தம்: 30 மணி நேரம்

பார்வை நூல்கள்

- தமிழர் நாகரிகமும் பண்பாடும், டாக்டர் அ. தட்சிணாமூர்த்தி, ஐந்திணைப் பதிப்பகம், 2001.
- 2. **தவறின்றித் தமிழ் எழுதுவோம்**, மா. நன்னன், ஏகம் பதிப்பகம், 1999.
- தவறின்றித் தமிழ் எழுத மருதார் அரங்கராசன், ஐந்திணைப் பதிப்பகம், 2003.
- தமிழ் இலக்கிய வரலாறு, வரதராசன், மு., புது தில்லி : சாகித்திய அக்காதெமி, 2002.
- புதிய தமிழ் இலக்கிய வரலாறு, நீல. பத்மநாபன், சிற்பி பாலசுப்ரமணியம், சாகித்திய அகாடெமி, 2007.
- செம்மொழி தமிழின் சிறப்பியல்புகள் முனைவர் மறைமலை இலக்குவனார்; <u>https://www.youtube.com/watch?v=HHZnmJb4jSY</u>
- 7. பாடநால் தேடலுக்கான இணையம் https://archive.org/

24LHIN21

HINDI

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वर्ष I – सत्र II (गद्य, पत्र लेखन & व्यावहारिक हिन्दी)

I Year-Sem II (Prose, Official Letter Writing & Functional Hindi)

Course Objectives :

- To inculcate the human values, importance of patriotism and hard work
- To train students in functional Hindi
- To introduce the usage of Inscript keyboard

UNIT I : मुंशी प्रेमचंद कृत 'बूढी काकी' (कहानी) 'Boodee kaki" (Kahani) by Munshi Premchand

UNIT II : जयशंकर प्रसाद कृत 'पुरस्कार' (कहानी) 'Puraskar' (Kahani) by Jaishankar Prasad

UNIT III: हरिशंकर परसाई कृत 'मैं नरक से बोल रहा हूँ' (व्यंग्य) 'Main Narak Se Bhol Raha Hun' (Vyangy) by Harishankar Parsayi,

UNIT IV: व्यावहारिक हिन्दी 1 – 50 – तकनीकि शब्द, 50 – पदनाम व विभागीय नाम, भाव एक भाषा अनेक Functional Hindi 1 - 50-Technical Words, 50-Designation & Department Names, Bhav Ek Bhasha Anek

UNIT V : व्यावहारिक हिन्दी 2 – पत्र लेखन – 3 औपचारिक पत्र, इन्स्क्रिप्ट कीबोर्ड का परिचय Functional Hindi 2 -Letter Writing- 3 Official Letters. Introduction to Inscript Keyboard

Total: 30hrs

6hrs.

6hrs.

6hrs.

6hrs

6hrs.

Course Outcome:

At the end of this course Students will be able to

- CO1 Know to the human values
- CO2 Know the importance of patriotism
- CO3 Know the value of hardwork in human life
- CO4 Journalise in Functional Hindi
- CO5 Use inscript keyboard

Text Book:

Ed. Subhash chandar, Boodi Kaki by Premchand (2012) Natioonal Book Trust, Jaishankar Prasad, Pratinidhi Kahaniyan, (2015) Raj Kamal Prakashan, Harishankar Parsai, Pratinidhi vyangy, (2007) Rajkamal. Kendriya Hindi Prashikshan Sansthan, Parangat, (2015) Bharat Sarkar.

Reference book:

Kendriya Hindi Sansthan, (2012) Banking Hindi Patyakram,

Weblink:

Munshi Premchand, Manasarovar, 2007, http://gadyakosh.org Jaishankar Prasad/ http://gadyakosh.org Harishankar Parsai/ https://hindikahani.hindi-kavita.com Prayojanmoolak Hindi:https://hi.mwikipedia.org 15 209 https://rajbhasha.gov.in/en/introduction

VEEDG

24LFRE21	FRENCH PAPER II	L T P O C
Course Objectives:		2 0 0 1 2
The lessons are being chosen:		
1) to express his / her where abo	outs and to ask seek direction	
2) to express obligation and rest		
3) to describe a place		
4) to narrate and to question		
5) to describe someone		
6) to express his desire and to sp	beak about the futur	
Units:		
1) C'est où	V. I CARLEN	
,	s au, à la…, Le passé composé et l'accord du par	ticiple 5 hours
2) N'oubliez pas Le pronom relatif Qui, que, où, Lo	s pronoms compléments indirects (me, te, lui, le	ur) 5 hours
3) Belle vue sur la mer -		
Les adjectifs démonstratifs, Y-pro	nom complément.	4 hours
4) Quel beau voyage!		
Les verbes pronominaux, En- pro	nom complément.	4 hours
5) Oh ! joli		
L'imparfait, L'imparfait ou le pass	é composé.	5 hours
6) Et après ?		
Le futur simple, Le subjonctif prés	ent.	7 hours
T 1 11 101		
Total no. of hours - 30 hours		- Co
Course Outcome:		18.3 × 1
	express his/her where about and to ask dire	ction
	express obligation and restriction	
3) The students would be able to		
4) The students would be able to		
5) The students would be able to		41.4
o) The students would be able to	express his desire and to speak about the fu	lui

<u>Text / Reference Book:</u> Prescribed book: LATITUDES 1 (A1/A2) MÉTHODE DE FRANÇAIS - Régine Mérieux and Yves Loiseau Reference book: SAISON A1 - MÉTHODE DE FRANÇAIS - Marie-Noëlle Cocton, Élodie Heu, Catherine Houssa, Émilie Kasazian

0 Р С Т **24LENG21 ENGLISH – II** 2 0 0 1 2 **Course Objectives:** • To read and understand different types of prose, poetry, and fiction. • To think critically about texts and express ideas clearly. • To recognize and discuss key themes and styles in literary works. • To learn and use grammar rules correctly in writing and speaking. • To write more effectively by applying grammar and literary techniques. Credit Hours **Unit I - Prose** 6 • If you are wrong, admit it- Dale Garnegie • Words of Wisdom- Chetan Bhaghat **Unit II - Poetry** 6 • La Belle Dame Sans Merci - John Keats • Ozymandias- P.B.Shelley **Unit III – Fiction** 6 • The School for Empathy - E.V. Lucas • The Lamb to the Slaughter-Roald Dahl **Unit IV - Grammar** 6 • Types of sentences, Concord **Unit V - Grammar** • Tenses, Voices Total **30 hours Course Outcomes:** At the end of this course, the students would have learnt to **CO1** Identify poetic expressions in the course of daily speech **CO2** Students will develop skills that enable them to communicate effectively in writing.

CO5 Strengthen the ability to solve life's problems, as highlighted in the selections.

writing.

References:

CO3

CO4

• Dr. M. Narayana Rao and Dr. B. G. Barki–Anu's Current English for Communication(AnuChitra). June 2012.

Discriminate against different sensibilities in approaching life.

• General English for competitive examinations by V.Rajagopalan (McGraw Hill Education) -2010.

Students will develop skills that enable them to communicate effectively in

24CBMB21 MICROBIAL GENETICS AND MOLECULAR BIOLOGY

COURSE OBJECTIVE:

The candidate will gain knowledge about the structure, shape, and significance of DNA and RNA. They will understand the synthesis of RNA and proteins along with their regulation. The role of genes as the basic units of expression will be explored in depth.

UNIT- I NUCLEIC ACIDS

DNA as genetic material (Griffith, Avery, Hershey and Chase experiments). Nucleic acids – definition and structure. Nucleoside, nucleotide: definition and structure. DNA & RNA: Double helical structure. A-DNA, B-DNA & Z-DNA (structure and differences). General structure and types of RNA.

UNIT-II REPLICATION

Conservative, dispersive, semi-conservative, bidirectional and semi-discontinuous, Theta model of replication, Mechanism of replication-Rolling-circle model. Prokaryotic DNA replication. Enzymes and proteins associated with DNA replication. Eukaryotic DNA replication.

UNIT-III TRANSCRIPTION

Transcription–general principles, basic apparatus, RNA polymerases and steps involved. Monocistronic and polycistronic mRNAs. Processing of RNA. Genetic code. Translation overview – Prokaryotes and Eukaryotes. Post translational modifications.

UNIT – IV GENE TRANSFER

Conjugation: Discovery, F+, F- and Hfr cells and F- genetic crosses. Transformation – competent cells – mechanism, transduction – generalized and specialized. Transposition Regulation of gene expression – structural and functional gene, operon – inducible operon – lac operon, repression operon – Trp operon, attenuation. Ara aperon.

UNIT-V MUTATION

Mutation and genetic analysis of mutants: Mutation and its types – insertion, deletion, addition, rearrangement. Genetic analysis. Mutagenesis: Types: Site directed mutagenesis, base analogue mutants, tautomerization. Physical and chemical mutagens.

Total: 45 Hours

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Course Outcome:

At the end of this course students will be able to:

CO1: Describe the flow of information from DNA to Protein and the three well-known mechanisms by which genetic material is transferred among microorganisms.

CO2: Explain different types of DNA replication and grasp the replication of singlestranded DNA molecules and the various features of retrovirus replication.

CO3: Compare the various cellular mechanisms involved in the control of prokaryotic and eukaryotic transcription.

CO4: Illustrate the mechanisms of gene transfer through conjugation, transformation, and transduction.

CO5: Identify different types of extrachromosomal elements; the nature of the transposable elements in the prokaryotic and eukaryotic cells.

CO6: Analyze the various mechanisms of genetic exchange, mutations, and their implications.

Text Books:

1. D. Freifelder (2008), Molecular Biology. Narosa Publishing House, New Delhi. *Reference Books:*

- S.R. Maloy, J.E. Cronan, D. Freifelder (1994), Microbial Genetics. Jones and Barlette publishers.
- H. Lodish, O. Baltimore, A, Berk S.L. Zipursky, P. Matsudaira, J. Darnell (1995) Molecular Cell Biology. Scientific American Books.
- 3. B. Lewin (2004) Genes VIII, Oxford University Press.
- William Haynes (1985), The Genetics of Bacteria and Their Viruses. Black well Scientific Publishers, Oxford.
- 5. P.B. Kaufman, W. Wu. D. Kim and L.J. Cseke, (2011) Molecular and Cellular Methods in Biology and Medicine, CRC Press, Florida.

Web Sources:

- https://bio.libretexts.org/Bookshelves/Genetics/Genetics_Agriculture_and_Biotechnology_ Suza_and_Lee)/01%3A_Chapters/1.11%3A_Recombinant_DNA_Technology
- 2. https://www.britannica.com/science/recombinant-DNA-technology
- 3. https://microbenotes.com/recombinant-dna-technology-steps-applications-and-limitations/

24CBMB22	INDUSTRIAL MICROBIOLOGY

COURSE OBJECTIVE:

The candidate will gain insight into the significance of microbes in the production of industrially important products. They will also learn about types of bioreactors and product separation technologies. Study microbial production of diverse industrial products like antibiotics, organic acids, enzymes, and beverages.

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UNIT-I INTRODUCTION OF INDUSTRIAL MICROBIOLOGY 9 Overview of industrial microbiology and its significance in various industries. Historical perspective: milestones and developments in industrial microbiology. Component parts of a fermentation process. Isolation, screening, improvement, preservation and handling the microbial strains.

UNIT-II INDUSTRIAL FERMENTATION MEDIA

Formulation strategies, economical means of providing energy, carbon, nitrogen, vitamin and mineral sources. Role of additional ingredients - buffers, precursors, chelators, inhibitors, inducers and antifoams. Sterilization of industrial fermentation media.

UNIT-III UPSTREAM PROCESSING

Types of fermentation processes - batch, fed-batch, continuous fermentations. Solid state fermentation and submerged fermentation. Components of a typical bio-reactor, Types of bioreactors- constantly stirred tank and air-lift fermenters, Measurement and control of fermentation parameters - pH, temperature, dissolved oxygen, foaming and aeration.

UNIT – IV DOWNSTREAM PROCESSING

Cell disruption, filtration, centrifugation, solvent extraction, precipitation, lyophilization and spray drying, Enzyme immobilization- Methods of immobilization, advantages and applications of immobilization, large scale applications of immobilized enzymes.

UNIT-V MICROBIAL PRODUCTION OF INDUSTRIAL PRODUCTS

Microbial production of: chemotherapeutic agents - penicillin, streptomycin, tetracycline; Organic acids- Citric acid, gluconic acid; Amino acids- L-Glutamic acid, L- Tryptophan, L-Lysine; Enzymes-amylase, protease, lipase. Production of Wine, beer, ethanol and Vitamin B12.

Total: 45 Hours

Course Outcome:

At the end of this course students will be able to:

CO1: Describe the principles governing microbial involvement in industrial processes.

CO2: Design and optimize fermentation media for efficient production of desired products using formulation strategies.

CO3: Operate various fermentation processes and bioreactor systems and its controlling key parameters.

CO4: Execute downstream processing techniques to extract, purify, and immobilize industrial products, demonstrating mastery in handling microbial cultures.

CO5: Evaluate the economic and environmental implications of microbial production processes, and propose optimizations for sustainable industrial practices.

CO6: Develop practical skills for handling microbial cultures, designing fermentation processes, and optimizing production techniques for industrial applications.

CO7: Analyze the genetic and metabolic pathways of microorganisms to enhance their productivity and stability in industrial applications.

Text Books:

1. A.H. Patel (1996), Industrial Microbiology, 1st edition, Macmillan India Limited

<u>Reference Books:</u>

- 1. N. Okafor (2007), Modern Industrial Microbiology and Biotechnology,1st edition, Bios Scientific Publishers Limited. USA.
- 2. M.J. Waites, N.L. Morgan, J.S. Rockey, and G. Higton (2001), Industrial Microbiology: An Introduction, 1st edition, Wiley – Blackwell.
- 3. A.N. Glaze and H. Nikaido (1995), Microbial Biotechnology: Fundamentals of Applied Microbiology, 1st edition, W.H. Freeman and Company.
- 4. L.E. Casida (1991), Industrial Microbiology, 1st edition, Wiley Eastern Limited.
- 5 W Crueger and A. Crueger (2000), Biotechnology: A textbook of Industrial Microbiology. 2nd edition, Panima Publishing Co. New Delhi.
- 6. P.F. Stanbury, A Whitaker and S.J. Hall (2006), Principles of Fermentation Technology, 2nd Edition, Wiley-Eastern.

Web Sources:

- https://bio.libretexts.org/Bookshelves/Microbiology/Microbiology_(Boundless)/17%3A_ Industrial_Microbiology
- 2. https://microbiologyclass.net/introduction-to-industrial-microbiology/#google_vignette
- 3. https://staff.run.edu.ng/oermedia/422231995398.pdf

24MBMB21

BIOINSTRUMENTATION

COURSE OBJECTIVE:

Overall, a bioinstrumentation course is designed to equip students with theoretical knowledge, practical skills, and critical thinking abilities necessary for careers in biomedical engineering, healthcare technology, and related fields. These objectives ensure that students can effectively contribute to the development, implementation, and improvement of biomedical instruments and systems aimed at enhancing healthcare delivery and patient outcomes.

UNIT- I BASIC INSTRUMENTATION AND BUFFER PREPARATION

Buffers - Preparation of Buffers – Standard Buffers – Molar and Normal Solutions PH - PH meter, Principles and applications of Autoclave – Hot air oven – Incubator, Laminar air flow chamber / Biosafety cabinets, BOD Incubator, Lyophilizer

UNIT-II CHROMATOGRAPHY

Partition principle, partition coefficient, Nature of partition forces. Adsorption Chromatography - Methods of paper chromatography, Thin layer chromatography and their Applications. Column Chromatography - Basic principle, method and application of - Gel filtration, Ion-exchange, and Affinity chromatography.

UNIT-III BIOMOLECULES

PCR-principles-instrumentation-applications- primer design- Nucleic acid sequencing methods-direct PCR sequencing. Protein estimation UV-lowry method- Bradford-purification methods- cell disruption- crude extract-fractionation methods. Enzyme assays-spectrophotometric and manometric methods. Immobilization of enzymes- physical and chemical methods.

UNIT – IV ELECTROPHORETIC TECHNIQUES

Electrophoretic techniques-principles-Electrophoresis of proteins-SDS-PAGE- Native gels-Gradient gels-Isoelectric focusing gels-Two dimensional PAGE-Cellulose acetate electrophoresis. Blotting technique-Southern blotting, Northern blotting, Western blotting, Electrophoresis of Nucleic acids-Agarose gel Pulse–field gel and Capillary electrophoresis. Chromatographic techniques, principles–materials and applications.

UNIT-V SPECTROPHOTOMETRY

Concept of electromagnetic radiation, spectrum of light, Concept of chromophores, extinction coefficient, Beer's law and its deviations, Difference between spectrophotometer and colorimeter. Basic principle, Instrumentation and Application - UV and visible

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Spectrophotometer, FTIR, IR Spectrophotometer and Mass spectrophotometer, NMR, Fluorescence.

Total: 45 Hours

Course Outcome:

At the end of this course students will be able to:

CO1: Demonstrate understanding of the principles and components of bioinstrumentation systems.

CO2: Operate and troubleshoot common bioinstrumentation devices such as biosensors, spectroscopy instruments, and imaging systems.

CO3: Apply calibration and validation procedures to ensure accuracy and reliability of bioinstrumentation measurements.

CO4: Interpret experimental results to draw conclusions about biological phenomena or conditions being studied.

CO5: Understand the interface between bioinstrumentation and biological systems.

CO6: Apply bioinstrumentation techniques to study physiological parameters, biomolecular interactions, or cellular processes.

CO7: Demonstrate awareness of regulatory requirements and standards relevant to bioinstrumentation development and usage.

Text Books:

- J. Jayaraman (2011), Laboratory Manual in Biochemistry, 2 nd Edition, Wiley Eastern Ltd., New Delhi.
- 2. P. Ponmurugan and P.B Gangathara (2012), Biotechniques, 1stEdition, MJP publishers.
- 3. L. Veerakumari (2009), Bioinstrumentation- 5th Edition, MJP publishers.
- Upadhyay, Upadhyay and Nath (2002), Biophysical chemistry Principles and techniques3rd Edition, Himalaya publishing home.

<u>Reference Books:</u>

- 1. Rodney.F.Boyer (2000), Modern Experimental Biochemistry, 3rd Edition, Pearson Publication.
- A. Skoog, M. West (2014), Principles of Instrumental Analysis,14th Edition, W.B.Saunders Co. Philadephia.
- 3. N.Gurumani. (2006), Research Methodology for biological sciences, 1 st Edition, MJP Publishers.
- 4. J.G. Webster (2004), Bioinstrumentation, 4 th Edition, John Wiley & Sons (Asia) Pvt.Ltd, Singapore.

24DBMB21

BASICS OF BIOCHEMISTRY

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4	0	0	2	4

COURSE OBJECTIVES

The candidate will gain knowledge about the structure, properties and functions of carbohydrates, proteins, lipids and nucleic acids. Basic biochemical techniques are also dealt with chromatography, centrifugation, and electrophoresis, providing a comprehensive understanding of essential metabolic pathways and analytical methods in biochemistry.

UNIT I CARBOHYDATES

Carbohydrates – Classification, structure of mono, oligo and polysaccharides. Glycolysis, TCA cycle. Disorders of Carbohydrate Metabolism: Diabetes mellitus.

UNIT II PROTEINS

Protein – Classification, amino acid – structure and classification. Biological role of proteins and structural organization of protein. Transamination, deamination and urea cycle and its regulation. Disorders of amino acid metabolism.

UNIT III LIPIDS

Lipids – Classification, Saturated and Unsaturated fatty acids. Biological functions of lipids. Biosynthesis and oxidation of fatty acids. Disorders of lipid metabolism.

UNIT IV NUCLEIC ACIDS

Nucleic acid – Nucleosides, Nucleotides, Structure of DNA and RNA – various types of RNA. Biosynthesis of purine and pyrimidine. Disorders of nucleic acid metabolism.

UNIT V CHROMATOGRAPHY

Chromatography – gel filtration, affinity, HPLC. Centrifugation – differential centrifugation. Electrophoresis – SDS-PAGE, Agarose gel electrophoresis.

Total: 60hrs

Course Outcome

At the end of the course, learners will be able to:

CO1: Recall and identify the structure and classification of carbohydrates, proteins, lipids, and nucleic acids.

CO2: Explain the metabolic pathways such as glycolysis, TCA cycle, transamination, deamination, the urea cycle, and fatty acid metabolism.

CO3: Apply biochemical techniques like chromatography, centrifugation, and electrophoresis to separate and analyze biomolecules.

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CO4: Analyze the regulation of biochemical processes and the interconnectivity of metabolic pathways in various disorders.

CO5: Design biochemical experiments to investigate the functions and interactions of biomolecules and interpret experimental data.

CO6: Evaluate the disorders of metabolism by means of various case study.

CO7: Gain the knowledge on different chromatographic methods.

Textbook:

1. J.L. Jain (2006), Fundamentals of Biochemistry, Chand Publications.

<u>Reference books:</u>

- 1. K. Robert (2005), Harper's Biochemistry, 26th edition, Murray Lance International Publication.
- 2. M.N. Chatterjee (2006), Text Book of Medical Biochemistry, 6th edition, Jaypee Publication.
- 3. U. Sathyanarayana (2006), Biochemistry, 3rd edition, Books and Allied (P) Ltd.



24DBMB22 BIOSAFTETY AND INTELLECTUAL PROPERTY RIGHTS (IPR)

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4	0	0	2	4

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COURSE OBJECTIVE:

The course aims to provide a comprehensive understanding of biosafety and risk assessment for handling biological agents safely. Familiarize with national and international biosafety regulations and the role of organizations like WHO and CDC. Explore the fundamentals of intellectual property rights (IPR) and their significance in biotechnology.

UNIT- I BIOSAFETY: INTRODUCTION AND GUIDELINES

Introduction, biosafety issues; Biological Safety Cabinets & their types; Primary Containment for Biohazards; Biosafety Levels of Specific Microorganisms. Biosafety Guidelines: Biosafety guidelines and regulations (National and International); GMOs/LMOs- Concerns and Challenges; Role of Institutional Biosafety Committees (IBSC), RCGM, GEAC etc. for GMO applications in food and agriculture;

UNIT-II RISK ANALYSIS AND GUIDELINES

Environmental release of GMOs; Risk Analysis; Risk Assessment; Risk management and communication; Overview of International Agreements - Cartagena Protocol. Use of Animals in Research and Testing, and Alternatives for Animals in Research, Animal Cloning, Human Cloning and their Ethical Aspects. Testing ofDrugs on Human Volunteers Public and Non-Governmental Organizations (NGOs)Participation in Biosafety and Protection of Biodiversity.

UNIT-III INTRODUCTION TO INTELLECTUAL PROPERTY

Introduction to Intellectual Property and History. Patents, Trademarks, Copyright, Trade secrets, Industrial Design and Rights, Traditional Knowledge, Geographical Indications - importance of IPR – patentable and non patentable – patenting life – legal protection of biotechnological inventions – World Intellectual Property Rights Organization (WIPO), Ethics, Pros and Cons of IP protection.

UNIT – IV GRANT OF PATENT, PATENTING AUTHORITIES AND TREATIES 12 Types of patent applications: Ordinary, PCT, Conventional, Divisional and Patent of Addition; An introduction to Patent Filing Procedures; Patent licensing and agreement; Patent infringement- meaning, scope, litigation, case studies, Rights and Duties of patent owner. Agreements and Treaties: GATT, TRIPS Agreements; WIPO Treaties; Budapest Treaty on international recognition of the deposit of microorganisms; UPOV & Brene conventions;

UNIT-V BIOSAFETY IN BIOTECHNOLOGY AND ETHICAL

CONSIDERATIONS

Definition and importance of biosafety in biotechnological research. Types of Biological Risks. Pathogenic microorganisms, genetically modified organisms (GMOs), and biohazards. Risk assessment strategies in biotechnology.

Total: 60 Hours

Course Outcome:

At the end of the course, learners will be able to:

CO1: Demonstrate a thorough understanding of the principles and practices of biosafety in biotechnology against the biological risks.

CO2: Analyze the ethical implications of using genetically modified organisms (GMOs) and other biotechnological innovations, considering their impact on human health and the environment.

CO3: Critically assess case studies related to biosafety incidents and improvements to current biosafety practices and regulations.

CO4: Develop effective communication strategies to engage with the public and address concerns regarding biotechnology and its implications for health and safety.

CO5: Explain the regulatory and ethical frameworks governing biotechnology, including national and international guidelines.

CO6: Collaborate effectively with peers and professionals from various disciplines to address biosafety challenges and ethical considerations in biotechnological research.

CO7: Evaluate emerging trends in biotechnology and their potential impact on biosafety regulations, ethical standards, and public policy.

Text Books:

4. . Shomini Parashar, Deepa Goel (2013), IPR, Biosafety and Bioethics Pearson India

Reference Books:

- R. J. M. A. Van den Borne, J. A. H. Van der Linde, and G. M. J. L. Thewessen (2018), Biosafety in Microbiological and Biomedical Laboratories, 5th edn, U.S. Department of Health and Human Services, Washington, D.C.
- B. J. Campbell, L. L. K. Trivett, and C. E. Redman (2017), Biotechnology and Biosafety: A Practical Guide, 1st edn, Wiley-Blackwell, Hoboken, NJ.

24PBMB21

PRACTICAL II

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PRACTICAL IN MOLECULAR BIOLOGY

COURSE OBJECTIVE:

The candidate will gain hands-on knowledge and acquire adequate skill required to separate and observe chromosomal DNA, RNA, amino acids, lipids as well as estimate nucleic acids.

- 1. Isolation of Antibiotic Resistant bacteria from soil samples.
- 2. Isolation of plasmid DNA and its demonstration by agarose gel electrophoresis.
- 3. Study survival curve of bacteria after exposure to ultraviolet (UV) light
- 4. Study the effect of chemical and physical (UV) mutagens on bacterial cells
- 5. Isolation of plasmid DNA and its demonstration by agarose gel electrophoresis.
- 6. Isolation of bacterial chromosomal DNA and demonstration.
- Quantitation of DNA and RNA by chemical methods-Dinitrophenol, orcinol, physical method – UV adsorption (Estimation of DNA by chemical method).
- 8. Estimation of proteins Lowry method; Bradford method
- 9. Electrophoretic methods PAGE native PAGE.
- 10. Separation of amino acids by paper chromatography.
- TLC Plant pigments, amino acids, lipids and vitamins. Protein separation by aqueous two phase partitioning.

Total: 45 Hours

Course Outcome:

At the end of this course students will be able to:

CO1: Explain the principles of chromatography and electrophoresis techniques.

CO2: Perform gel electrophoresis and HPLC experiments using proper techniques and safety protocols.

CO3: Analyze experimental data from molecular separation experiments.

CO4: Evaluate the reliability of chromatographic separations for different types of molecules.CO5: Design a protocol for a novel molecular separation experiment to separate specific

biomolecules.

CO6: Design a protocol for a novel molecular separation experiment to address a specific research question or problem.

CO7: List different types of molecular separation methods and their basic components.

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PRACTICAL III

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PRACTICAL IN INDUSTRIAL MICROBIOLOGY COURSE OBJECTIVE:

Develop skills in isolating and identifying soil bacteria producing amylase, protease, lipase, organic acids, and antibiotics; understand their industrial applications. Learn about fermenter design, components, sterilization, and control mechanisms; observe industrial fermenters and downstream processing. Master techniques for yeast immobilization, isolate lactic acid bacteria from curd, and produce wine, gaining insights into microbial roles and biochemical changes in fermentation processes.

- 1. Study of the different parts of fermenter
- 2. Screening for amylase producing bacteria from soil.
- 3. Screening for protease producing bacteria from soil.
- 4. Screening for lipase producing bacteria from soil.
- 5. Screening for organic acid producers (acetic acid/lactic acid)
- 6. Screening for antibiotic producers.
- 7. Immobilization of yeast.
- 8. Isolation of lactic acid bacteria from curd.
- 9. Wine Production.
- 10. A visit to any industry to see an industrial fermenter, and other downstream processing operations.

Total: 45 Hours

Course Outcome:

At the end of the course, learners will be able to:

CO1: Identify the different components of a fermenter and their respective roles in the fermentation process.

CO2: Employ methods to identify microorganisms capable of producing specific enzymes such as amylase and protease, and understand the principles behind these screening processes.

CO3: Screen microorganisms for their ability to produce organic acids such as acetic acid and lactic acid, and interpret results to select potential industrial strains.

CO4: Conduct assays to identify microorganisms with antibiotic-producing capabilities.

CO5: Understand various methods used for immobilizing yeast cells and their applications in industrial processes, and be able to perform basic immobilization procedures.

CO6: Analyze data obtained from fermentation experiments to optimize production conditions and improve yield.

CO7: Gain practical insights into industrial fermentation and downstream processing through industry visits.

Reference Books:

- 1. T.C.K. Sugitha, P. Raja, R. Rajesh, U. Sivakumar (2019), Practical manual cum Work Book "Industrial Microbiology"
- 2. Basanta Kumar Rai, Dil Kumar Subba (2016), Basic Practical Manual on Industrial Microbiology First Edition.

Web Sources:

1. file:///C:/Users/Suresh/Downloads/IMPracticalManual2020%20(1).pdf



24DVAC21

COMMUNICATION SKILLS

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Course Objectives:

- To develop effective verbal and non-verbal communication techniques for variouscontexts.
- To enhance listening skills for better comprehension and engagement in conversations.
- To improve written communication abilities, focusing on clarity, coherence, and style.
- To build confidence in public speaking through practice and constructive feedback.
- To cultivate interpersonal skills for successful collaboration and professional interactions.

Credit Hours

6

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Unit I –Introduction to Communication Skills

- Fundamentals of Communications
- Elements of Communication, Types of Communication

Unit II - Practical English

- Importance of the language Word Usage and Jargon
- Tenses and the effectiveness Basics of grammar (Noun/Verb/Adverb/Conjunction)

Unit III - Effective Communication

- LSRW (Listening, Speaking, Reading & Writing)
- Pronunciation Vocabulary Building
- Intonations & its importance

Unit IV - Workplace Communication

- Basics of telephone etiquette
- E-Mail writing
- Presentation Skills
- Interpersonal Skills
- Business English

Unit V - Quantitative Ability

- Verbal Ability Verbal Analogy
- Debating Skills Public Speaking

Total

30 Hours

Course Outcomes:

At the end of this course, the students would have learnt to

CO1	enhance participants' business communication skills	
CO2	enhance LSRW Skills (LSRW – Listening, Speaking, Reading & Writing)	
CO3	express opinions at free will in social/ personal gathering	
CO4	impact leadership qualities among participants	
CO5	engage in conversation with others to exchange ideas	

References:

- Basic communication skills for Technology, Andreja. J. Ruther Ford, 2nd Edition, Pearson Education, 2011
- Personality development and soft skills, Barun K Mitra, 1 stEdition, Oxford Press, 2011
- Elizabeth Harren, 7 April 2022, last updated: 16 November, 2023
- Kerry Patterson, Joseph Grenny, Ron McMillan, Al Switzler (McGraw-Hill)
- Ethan Beute and Stephen Pacinelli (Greenleaf)

NOWLEDG

• Soft skills and professional communication, Francis Peters SJ, 1stEdition, McGraw Hill Education, 2011

SOFT SKILLS II

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Credit Hours

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24SSKU21

Course Objectives:

- To develop strategies to enhance teamwork and collaboration in professional settings.
- To cultivate a positive attitude and mindset to foster constructive relationships and productivity.
- To develop leadership, decision-making and team bonding skills

Unit I - PROFESSIONAL BEHAVIOUR

- Team Building Team Bonding
- Inter-Personal Relationship-Intra-Personal Relationship

Unit II - PERSONALITY DEVELOPMENT

- Types of Personality
- Self-Confidence Confidence Building
- Attitude (Positive/Negative)

Unit III - TELEPHONE ETIQUETTE

- Basics of telephone etiquette
- Giving clear and concise information
- Tone & rate of speech
- Intonations & its Importance
- Whatsapp Communications

Unit IV - DECISION MAKING

- Types of Decisions planned-unplanned, individual-group, major-minor
- Types of Leadership styles Autocratic, democratic, lesse-faire, participative, beurocratic

Unit V - PROFESSIONAL ETIQUETTE

- Respect Salutations
- Official Behaviour

Total

30 Hours

6

Course Outcomes:

At the end of this course, the students will be able to,

CO1	understand the principles of effective team building and apply strategies to foster team bonding and cohesion in professional settings.
CO2	become self-confident individuals by mastering interpersonal skills, team management skills, and leadership skills.
CO3	practice techniques for effective communication in telephone conversations.
CO4	evaluate decision-making processes and their implications in professional settings.
CO5	exhibit professional conduct and demeanour in various professional situations.

<u>References:</u>

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- Nelson-Jones, R. (1992). Life skills, a handbook, Trowbridge, Wilts: Detesios Ltd.